PATENT COOPERATION TREATY

From the INTERNATIONAL BUREAU

PCT

NOTIFICATION OF ELECTION

(PCT Rule 61.2)

To:

Assistant Commissioner for Patents
United States Patent and Trademark Office
Box PCT
Washington, D.C.20231
ETATS-UNIS D'AMERIQUE

in its capacity as elected Office

Date of mailing (day/month/year) 11 August 2000 (11.08.00)	
International application No. PCT/EP99/10470	**Applicant's or agent's file reference** B4140A-GD
International filing date (day/month/year) 30 December 1999 (30.12.99)	**Priority date** (day/month/year) 30 December 1998 (30.12.98)
Applicant GIANNI, Alessandro, Massimo	

1. The designated Office is hereby notified of its election made:

   [X] in the demand filed with the International Preliminary Examining Authority on:

   03 July 2000 (03.07.00)

   [ ] in a notice effecting later election filed with the International Bureau on:

2. The election [X] was

   [ ] was not

   made before the expiration of 19 months from the priority date or, where Rule 32 applies, within the time limit under Rule 32.2(b).

The International Bureau of WIPO **34, chemin des Colombettes** **1211 Geneva 20, Switzerland**	Authorized officer Olivia RANAIVOJAONA
Facsimile No.: (41-22) 740.14.35	Telephone No.: (41-22) 338.83.38

PATENT COOPERATION TREATY

From the
INTERNATIONAL PRELIMINARY EXAMINING AUTHORITY

To:

ERNEST GUTMANN-YVES PLASSERAUD S.A.
3, rue Chauveau-Lagarde
F-75008 Paris
FRANCE

REÇU LE
16 MARS 2001
E. GUTMANN - Y. PLASSERAUD S.A.

PCT

NOTIFICATION OF TRANSMITTAL OF THE INTERNATIONAL PRELIMINARY EXAMINATION REPORT
(PCT Rule 71.1)

Date of mailing *(day/month/year)*	14.03.2001
Applicant's or agent's file reference B4140A-GD	**IMPORTANT NOTIFICATION**

International application No.	International filing date *(day/month/year)*	Priority date *(day/month/year)*
PCT/EP99/10470	30/12/1999	30/12/1998

Applicant
APPLIED RESEARCH SYSTEMS ARS HOLDING N.V. et al.

1. The applicant is hereby notified that this International Preliminary Examining Authority transmits herewith the international preliminary examination report and its annexes, if any, established on the international application.

2. A copy of the report and its annexes, if any, is being transmitted to the International Bureau for communication to all the elected Offices.

3. Where required by any of the elected Offices, the International Bureau will prepare an English translation of the report (but not of any annexes) and will transmit such translation to those Offices.

4. **REMINDER**

The applicant must enter the national phase before each elected Office by performing certain acts (filing translations and paying national fees) within 30 months from the priority date (or later in some Offices) (Article 39(1)) (see also the reminder sent by the International Bureau with Form PCT/IB/301).

Where a translation of the international application must be furnished to an elected Office, that translation must contain a translation of any annexes to the international preliminary examination report. It is the applicant's responsibility to prepare and furnish such translation directly to each elected Office concerned.

For further details on the applicable time limits and requirements of the elected Offices, see Volume II of the PCT Applicant's Guide.

Name and mailing address of the IPEA/	Authorized officer
European Patent Office D-80298 Munich Tel. +49 89 2399 - 0 Tx: 523656 epmu d Fax: +49 89 2399 - 4465	Hundt, D Tel.+49 89 2399-8042

Form PCT/IPEA/416 (July 1992)

PATENT COOPERATION TREATY

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INTERNATIONAL SEARCH REPORT

(PCT Article 18 and Rules 43 and 44)

Applicant's or agent's file reference	FOR FURTHER ACTION	see Notification of Transmittal of International Search Report (Form PCT/ISA/220) as well as, where applicable, item 5 below.
B4140A-GD		
International application No.	International filing date *(day/month/year)*	(Earliest) Priority Date *(day/month/year)*
PCT/EP 99/ 10470	30/12/1999	30/12/1998

Applicant

APPLIED RESEARCH SYSTEMS ARS HOLDING N.V. et al.

This International Search Report has been prepared by this International Searching Authority and is transmitted to the applicant according to Article 18. A copy is being transmitted to the International Bureau.

This International Search Report consists of a total of 5 sheets.

[X] It is also accompanied by a copy of each prior art document cited in this report.

1. **Basis of the report**

   a. With regard to the **language**, the international search was carried out on the basis of the international application in the language in which it was filed, unless otherwise indicated under this item.

   [ ] the international search was carried out on the basis of a translation of the international application furnished to this Authority (Rule 23.1(b)).

   b. With regard to any **nucleotide and/or amino acid sequence** disclosed in the international application, the international search was carried out on the basis of the sequence listing :

   [ ] contained in the international application in written form.

   [ ] filed together with the international application in computer readable form.

   [ ] furnished subsequently to this Authority in written form.

   [ ] furnished subsequently to this Authority in computer readble form.

   [ ] the statement that the subsequently furnished written sequence listing does not go beyond the disclosure in the international application as filed has been furnished.

   [ ] the statement that the information recorded in computer readable form is identical to the written sequence listing has been furnished

2. [X] **Certain claims were found unsearchable** (See Box I).

3. [ ] **Unity of invention is lacking** (see Box II).

4. With regard to the **title**,

   [X] the text is approved as submitted by the applicant.

   [ ] the text has been established by this Authority to read as follows:

5. With regard to the **abstract**,

   [X] the text is approved as submitted by the applicant.

   [ ] the text has been established, according to Rule 38.2(b), by this Authority as it appears in Box III. The applicant may, within one month from the date of mailing of this international search report, submit comments to this Authority.

6. The figure of the drawings to be published with the abstract is Figure No. _____

   [ ] as suggested by the applicant.

   [ ] because the applicant failed to suggest a figure.

   [ ] because this figure better characterizes the invention.

   [X] None of the figures.

REPLACED BY ART 34 AMDT

PATENT COOPERATION TREATY

REC'D 16 MAR 2001
WIPO PCT

PCT

14

INTERNATIONAL PRELIMINARY EXAMINATION REPORT

(PCT Article 36 and Rule 70)

Applicant's or agent's file reference	FOR FURTHER ACTION	See Notification of Transmittal of International Preliminary Examination Report (Form PCT/IPEA/416)
B4140A-GD		
International application No.	International filing date *(day/month/year)*	Priority date *(day/month/year)*
PCT/EP99/10470	30/12/1999	30/12/1998
International Patent Classification (IPC) or national classification and IPC A61K38/27		
Applicant APPLIED RESEARCH SYSTEMS ARS HOLDING N.V. et al.		

1. This international preliminary examination report has been prepared by this International Preliminary Examining Authority and is transmitted to the applicant according to Article 36.

2. This REPORT consists of a total of 10 sheets, including this cover sheet.

   ☒ This report is also accompanied by ANNEXES, i.e. sheets of the description, claims and/or drawings which have been amended and are the basis for this report and/or sheets containing rectifications made before this Authority (see Rule 70.16 and Section 607 of the Administrative Instructions under the PCT).

   These annexes consist of a total of 10 sheets.

3. This report contains indications relating to the following items:

   - I ☒ Basis of the report
   - II ☐ Priority
   - III ☒ Non-establishment of opinion with regard to novelty, inventive step and industrial applicability
   - IV ☐ Lack of unity of invention
   - V ☒ Reasoned statement under Article 35(2) with regard to novelty, inventive step or industrial applicability; citations and explanations suporting such statement
   - VI ☐ Certain documents cited
   - VII ☐ Certain defects in the international application
   - VIII ☒ Certain observations on the international application

Date of submission of the demand	Date of completion of this report
03/07/2000	14.03.2001
Name and mailing address of the international preliminary examining authority: European Patent Office D-80298 Munich Tel. +49 89 2399 - 0 Tx: 523656 epmu d Fax: +49 89 2399 - 4465	Authorized officer Fayos, C Telephone No. +49 89 2399 2180

Form PCT/IPEA/409 (cover sheet) (January 1994)

INTERNATIONAL PRELIMINARY EXAMINATION REPORT

International application No. PCT/EP99/10470

I. Basis of the report

1. This report has been drawn on the basis of *(substitute sheets which have been furnished to the receiving Office in response to an invitation under Article 14 are referred to in this report as "originally filed" and are not annexed to the report since they do not contain amendments (Rules 70.16 and 70.17).)*:
Description, pages:

1-65 as originally filed

Claims, No.:

1-55 with telefax of 10/11/2000

Drawings, sheets:

1/2,2/2 as originally filed

2. With regard to the **language**, all the elements marked above were available or furnished to this Authority in the language in which the international application was filed, unless otherwise indicated under this item.

These elements were available or furnished to this Authority in the following language: , which is:

- ☐ the language of a translation furnished for the purposes of the international search (under Rule 23.1(b)).
- ☐ the language of publication of the international application (under Rule 48.3(b)).
- ☐ the language of a translation furnished for the purposes of international preliminary examination (under Rule 55.2 and/or 55.3).

3. With regard to any **nucleotide and/or amino acid sequence** disclosed in the international application, the international preliminary examination was carried out on the basis of the sequence listing:

- ☐ contained in the international application in written form.
- ☐ filed together with the international application in computer readable form.
- ☐ furnished subsequently to this Authority in written form.
- ☐ furnished subsequently to this Authority in computer readable form.
- ☐ The statement that the subsequently furnished written sequence listing does not go beyond the disclosure in the international application as filed has been furnished.
- ☐ The statement that the information recorded in computer readable form is identical to the written sequence listing has been furnished.

4. The amendments have resulted in the cancellation of:

- ☐ the description, pages:
- ☐ the claims, Nos.:

☐ the drawings, sheets:

5. ☐ This report has been established as if (some of) the amendments had not been made, since they have been considered to go beyond the disclosure as filed (Rule 70.2(c)):

(Any replacement sheet containing such amendments must be referred to under item 1 and annexed to this report.)

6. Additional observations, if necessary:

III. Non-establishment of opinion with regard to novelty, inventive step and industrial applicability

1. The questions whether the claimed invention appears to be novel, to involve an inventive step (to be non-obvious), or to be industrially applicable have not been examined in respect of:

☐ the entire international application.

☒ claims Nos. 18 (completely), 1-17 and 19-28 (industrial applicability).

because:

☒ the said international application, or the said claims Nos. 1-17 and 19-28 (industrial applicability) relate to the following subject matter which does not require an international preliminary examination (*specify*):
see separate sheet

☐ the description, claims or drawings (*indicate particular elements below*) or said claims Nos. are so unclear that no meaningful opinion could be formed (*specify*):

☐ the claims, or said claims Nos. are so inadequately supported by the description that no meaningful opinion could be formed.

☒ no international search report has been established for the said claims Nos. 18 (completely).

2. A meaningful international preliminary examination report cannot be carried out due to the failure of the nucleotide and/or amino acid sequence listing to comply with the standard provided for in Annex C of the Administrative Instructions:

☐ the written form has not been furnished or does not comply with the standard.

☐ the computer readable form has not been furnished or does not comply with the standard.

V. Reasoned statement under Article 35(2) with regard to novelty, inventive step or industrial applicability; citations and explanations supporting such statement

1. Statement

Novelty (N) Yes: Claims 1-17 and 19-55

	No:	Claims	-
Inventive step (IS)	Yes:	Claims	1-17 and 19-55
	No:	Claims	-
Industrial applicability (IA)	Yes:	Claims	29-55; claims 1-17 and 19-28 see separate sheet
	No:	Claims	

2. Citations and explanations
see separate sheet

VIII. Certain observations on the international application

The following observations on the clarity of the claims, description, and drawings or on the question whether the claims are fully supported by the description, are made:
see separate sheet

Re Item III

Non-establishment of opinion with regard to novelty, inventive step and industrial applicability

1- Claims 1-28 relate to subject-matter considered by this Authority to be covered by the provisions of Rule 67.1(iv) PCT. Consequently, no opinion will be formulated with respect to the industrial applicability of the subject-matter of these claims (Article 34(4)(a)(i) PCT).

1.1- The amendment in claim 18: "or one of its derivatives or any factor inducing GH release" is not allowable because it goes beyond the disclosure of the international application as filed (Art. 34 (b) PCT).

Re Item V

Reasoned statement under Article 35(2) with regard to novelty, inventive step or industrial applicability; citations and explanations supporting such statement

2- Reference is made to the following documents:

D1: PATENT ABSTRACTS OF JAPAN vol. 095, no. 007, 31 August 1995 (1995-08-31) & JP 07 101877 A (MITSUI TOATSU CHEM INC), 18 April 1995 (1995-04-18)

D2: TIAN Z G ET AL: 'Recombinant human growth hormone promotes hematopoietic reconstitution after syngeneic bone marrow transplantation in mice.' STEM CELLS, (1998) 16 (3) 193-9., XP002114334

D3: BREGNI M ET AL: 'Comparative effects of granulocyte-macrophage colony-stimulating factor and granulocyte colony - stimulating factor after high-dose cyclophosphamide cancer therapy.' JOURNAL OF CLINICAL ONCOLOGY, (1996 FEB) 14 (2) 628-35. , XP000877223

D4: MURPHY W J ET AL: 'GROWTH HORMONE EXERTS HEMATOPOIETIC GROWTH-PROMOTING EFFECTS IN VIVO AND PARTIALLY COUNTERACTS THE MYELOSUPPRESSIVE EFFECTS OF AZIDOTHYMIDINE' BLOOD, vol. 80, no. 6, 15 September 1992 (1992-09-15),

pages 1443-1447, XP002065327

D5: MIYASHITA Y: 'The effect of growth hormone on leukopoiesis: in vivo and in vitro studies.' NIPPON NAIBUNPI GAKKAI ZASSHI. FOLIA ENDOCRINOLOGICA JAPONICA, (1991 JUL 20) 67 (7) 785-95., XP002114569

D6: MURPHY W J ET AL: 'Human growth hormone promotes engraftment of murine or human T cells in severe combined immunodeficient mice.' PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES OF THE UNITED STATES OF AMERICA, (1992 MAY 15) 89 (10) 4481-5., XP002114570

D7: US-A-5 199 942 (GILLIS STEVEN) 6 April 1993 (1993-04-06)

D8: KÖRBLING M ET AL: 'Allogeneic peripheral blood stem cell transplantation using normal patient-related pediatric donors' BONE MARROW TRANSPLANTATION, vol. 18, no. 5, November 1996 (1996-11), pages 885-890, XP002114335

D9: OHMIZONO Y ET AL: 'THROMBOPOIETIN AUGMENTS EX VIVO EXPANSION OF HUMAN CORD BLOOD -DERIVED HEMATOPOIETIC PROGENITORS IN COMBINATION WITH STEM CELL FACTOR AND FLT3 LIGAND' LEUKEMIA, vol. 11, no. 4, 1 April 1997 (1997-04-01), pages 524-530, XP002053902

D10: WO 94 28916 A (BRITISH BIOTECH PHARM ;COMER MICHAEL BERISFORD (GB); MCCOURT MATTH) 22 December 1994 (1994-12-22)

D11: US-A-5 649 904 (GIANNI ALESSANDRO M) 22 July 1997 (1997-07-22)

NOVELTY - Art. 33 (1) and (2) PCT

3- Claims 1-17 and 19-55 appear to be novel over the prior art cited in the search report.

3.1- D2 discloses the administration of rhGH (ip injection, 10 µg, every other day, total of 10 injections) to mice after syngeneic bone marrow transplantation (BMT) to determine its effect on hematopoietic reconstitution. D2 shows that mice that received rhGH exhibited significant increases in total hematopoietic progenitor cell

content in both bone marrow and spleen, erythroid cell progenitor content was also significantly increased and analysis of peripheral blood indicated that administration of rhGH resulted in significant increases in the rate of white blood cell, red blood cell, hematocrit and platelet recovery (abstract and p 197 c 1 - see also materials and methods). D2 proposes (discussion) the clinical use of rhGH in BMT, chemotherapy, anemia and neutropenia, and states (discussion) the possible combination with G-CSF and GM-CSF (possible synergistic effect).

[see assay for in vitro hematopoisesis p 194 c 1 § 4 - c 2 § 1]

D2 differs from the present application in that D2 does not disclose "circulating cells capable of regenerating hematopoiesis in vivo", since D2 discloses the increase in the levels of white blood cells, red blood cells, hematocrit (whole cell count in the blood) and platelets in the peripheral blood by administering growth hormone; these cells are differentiated, not stem cells, and are consequently not capable of regenerating hematopoiesis in vivo. Furthermore, D2 mentions that mice that received rhGH exhibited significant increases in total hematopoietic progenitor cell content (cfu) in both bone marrow and spleen. hence, D2 does not disclose at all the use of growth hormone to increase the number of circulating cells capable of regenerating hematopoiesis in vivo.

Hence, D2 does not appear to destroy the novelty of claim 3 and dependent claims 4, 8, 9 , 11, 21, 25, 26 and claims 32, 37, 42, 43, 44, 45, 48 and 49.

3.2- In D4 (abstract), BALB/c mice and mice with severe combined immune deficiency (SCID), which lack T cells and B cells, were administered intraperitoneal injections of rhGH for 7 days (20 µg ip injection every other day). Upon analysis, both strains of mice exhibited an increase in splenic and bone marrow hematopoietic progenitor cell content and cellularity, indicating that rhGH can act as a hematopoietic growth factor (see also materials and methods). D4 also shows that treatment of mice with rhGH partially counteracted the myelosuppressive properties of AZT. Bone marrow cellularity, hematocrit values, white blood cell counts, and splenic hematopoietic progenitor cell content were all significantly increased if rhGH was concurrently administered with AZT. Thus, rhGH exerts significant direct hematopoietic growth-promoting effects in vivo and may be of potential clinical use to promote

hematopoiesis in the face of myelotoxic therapy. Finally, D4 proposes rhGH for the treatment after BMT, chemotherapy, radiotherapy, anemia and neutropenia and suggests its use for augmenting hematopoiesis in humans.

[see assay for in vitro hematopoisesis p 1443 c 2 § 1]

Although D4 discloses the increase of bone marrow cellularity, hematocrit values, white blood cell counts and splenic hematopoietic progenitor cell content by human growth hormone, as mentioned above with regard to the hematocrit and white blood cell counts, these cells are not cells capable of regenerating hematopoiesis in vivo. With regard to the bone marrow cellularity and splenic hematopoietic progenitor cell content, again, these cells may comprise stem cells, but D4 does not report that their content has been increased in the blood circulation.

Hence, D4 does not appear to destroy the novelty of claim 3 and dependent claims 4, 8, 11, 21, 25, 26 and claims 32, 37, 42, 43, 44, 45 and 49.

3.3- D6 shows that rhGH (three 5 µg injections) promotes the peripheral engraftment of murine or human T-cells in severe combined immune deficiency SCID mice and proposes rhGH for clinical application in AIDS after BMT.

D6 does not disclose the use of growth hormone to prevent/treat opportunistic infections after transplantation or for limiting the risk of tumor recurrence after transplantation, and hence does not appear to destroy the novelty of claim 42.

4- Claims 1-2, 5-7, 10, 12-17, 19-20, 22-24, 27-31, 33-36, 38-41, 46-47 and 50-55 appear to be novel in the light of the available prior art.

4.1- The novel feature appears to be the isolation of produced hematopoietic cells from a donor for injection into an acceptor.

INVENTIVE STEP - Art. 33 (1) and (3) PCT

5- The present application is based on the finding that growth hormone increases the number of circulating cells capable of regenerating hematopoiesis in vivo, in other words that growth hormone possesses mobilisation properties.

This finding is neither explicitly disclosed, nor suggested in the prior art documents cited in the search report, alone or combined.

Claims 1-17 and 19-55 can therefore be considered as being inventive.

5.1- The problem posed in the present application is therefore to provide means for regenerating hematopoiesis in vivo.

The solution proposed in the present application is the preparation of a population of circulating cells capable of regenerating hematopoiesis in vivo in a donor, said cells being used to treat, by re-infusion, transplantation or engraftment a recipient in need of such cells

INDUSTRIAL APPLICABILITY - Art. 33 (1) and (4) PCT

6- For the assessment of the present claims 1-28 on the question whether they are industrially applicable, no unified criteria exist in the PCT Contracting States. The patentability can also be dependent upon the formulation of the claims. The EPO, for example, does not recognize as industrially applicable the subject-matter of claims to the use of a compound in medical treatment, but may allow, however, claims to a known compound for first use in medical treatment and the use of such a compound for the manufacture of a medicament for a new medical treatment.

6.1- Claims 29-55 appear to be industrially applicable.

Re Item VIII

Certain observations on the international application

7- Abbreviations should have been specified in the claims, since the claims should be self understanding.

8- The vague and imprecise terms: "around or more" in the claims implies that the subject-matter for which protection is sought may be different to that defined by the claims, thereby resulting in lack of clarity (Article 6 PCT).

9- Although claims 1, 2, 3 on the one hand and claims 27, 30, 31, 32 and 42 on the other hand, have been drafted as separate independent claims, they still appear to relate effectively to the same subject-matter and to differ from each other only with regard to the definition of the subject-matter for which protection is sought and/or in respect of the terminology used for the features of that subject-matter. The aforementioned claims therefore lack conciseness. Moreover, lack of clarity of the claims as a whole arises, since the plurality of independent claims makes it difficult, if not impossible, to determine the matter for which protection is sought, and places an undue burden on others seeking to establish the extent of the protection.

Hence, claims 1-55 do not meet the requirements of Article 6 PCT.

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WORLD INTELLECTUAL PROPERTY ORGANIZATION
International Bureau

INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification 7 :
A61K 38/27, C12N 5/06, 5/08, A61K 35/14, 35/28 // (A61K 38/27, 38:19)

A1

(11) International Publication Number: WO 00/40260

(43) International Publication Date: 13 July 2000 (13.07.00)

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98124834.7 30 December 1998 (30.12.98) EP

(71) Applicant *(for all designated States except US)*: APPLIED RESEARCH SYSTEMS ARS HOLDING N.V. [NL/NL]; Pietermaai 15, Curacao (AN).

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(74) Agents: GUTMANN, Ernest et al.; Ernest Gutmann–Yves Plasseraud S.A., 3, rue Chauveau–Lagarde, F–75008 Paris (FR).

(81) Designated States: AE, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CR, CU, CZ, DE, DK, DM, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).

Published
With international search report.

(54) Title: HUMAN GROWTH HORMONE TO STIMULATE MOBILIZATION OF PLURIPOTENT HEMATOPOIETIC STEM CELLS

(57) Abstract

The invention relates to the field of hematopoietic CD34+ cell mobilization. In particular, this invention relates to a method for increasing the mobilization of pluripotent hematopoetic CD34+ cells by administration of human growth hormone or one of its derivates to the individual.

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Human growth hormone to stimulate mobilization of pluripotent hematopoietic stem cells

Bone marrow transplantation (BMT) is a clinical procedure in which pluripotent hematopoietic cells obtained from bone marrow are transplanted to a patient. BMT is the treatment of choice in several hematological disorders, including malignancies, Severe Combined Immune Deficiencies (SCIDs), congenitally or genetically determined hematopoietic abnormalities, anemia, aplastic anemia, leukemia and osteopetrosis (Fischer et al., 1998). In the last ten years, the use of BMT grew from less than 5'000 to more than 40'000 annually (Waters et al., 1998).

Under steady state condition, the majority of hematopoietic stem and progenitor cells reside in the bone marrow and only a low number of these cells are detectable in peripheral blood. However, additional stem cells can be mobilized into the peripheral blood by treatment with myelosuppressive agents and/or certain hematopoietic growth factors (Van Hoef, 1998). Studies have demonstrated that peripheral blood stem cells (PBSC) infused in a host exhibits enhanced potential for engraftment as compared to bone marrow-derived stem and progenitor cells (Gianni et al., 1989 ; Larsson et al. , 1998). Thus, PBSC mobilized by chemotherapy, hematopoietic growth factors or the combination of these modalities are currently used in both autologous and nonautologous transplantation settings (Van Hoef, 1998 ; Anderlini and Korbling, 1997). In the case of nonautologous transplantation, the donors of stem cells are normal individuals and the procedure for mobilization of stem cells into the blood stream has to be achieve with minimal discomfort. In this case, stem cells

mobilization with hematopoietic growth factors is preferred to the treatment with antiblastic drugs (i.e. cyclophosphamid).

Several hematopoietic growth factors, such as G-CSF, EPO and CSF have been studied as mobilizing agents and are currently used to increase the number of PBSC prior to leukapheresis (Henry, 1997; Weaver and Testa, 1998). Treatments aimed at stimulating the overall hematopoiesis may be of great interest to mobilize a large set of progenitor cells and stem cells. Increased mobilization of stem cells is extremely valuable in the context of hematopoietic stem cells transplantation by reducing the number of leukapheresis required to collect sufficient amount of hematopoietic stem cells to be transplanted.

The first part of the invention provides a new mobilising agent used to increase the number of circulating cells capable of regenerating hematopoiesis in vivo in an individual.

The new mobilizing agent of the invention is growth hormone and especially Human Growth Hormone (hGH) or one of its derivatives or any factor inducing growth hormone release.

Unless it is otherwise specified, the term « GH » means Growth Hormone, one of its derivatives or any factor inducing growth hormone release within the context of the invention.

It has now been found that, by administering growth hormone and especially Human Growth Hormone (hGH) or one of its derivatives or any factor inducing growth hormone release, a mobilization of cells capable of regenerating hematopoiesis in vivo is obtained in the peripheral blood. Therefore, growth hormone and especially human Growth Hormone (hGH) or one of its derivatives or any factor inducing growth hormone release, administered

alone or in combination with other factors, represents a new method or use to mobilize cells capable of regenerating hematopoiesis in vivo to the peripheral blood.

Human Growth Hormone (hGH), also known as somatotropin is a protein hormone produced and secreted by the somatotropic cells of the anterior pituitary. hGH plays a key role in somatic growth through its effects on the metabolism of proteins, carbohydrates and lipids. In addition to its effects on somatic growth, hGH has been shown to stimulate blood cells in vitro (Derfalvi et al., 1998 ; Merchav et al; 1988), to increase erythrocytes and hemoglobin counts (Valerio et al. , 1997 ; Vihervuori et al. , 1996), to enhance both proliferation and Ig production in plasma cell lines (Kimata and Yoshida, 1994) and to stimulate $CD8^+$ cell counts and, to a lesser extent $CD4^+$ cell counts (Geffner, 1997).

The methods and uses of the invention which use the mobilising agent of the invention have several advantages :

- There is a low number of circulating cells capable of regenerating hematopoiesis. This number is considered insufficient to provide a cells engraftment dose by single or multiple apheresis in a reasonable time period. Methods and uses of the invention solve this problem by a temporary peripheralization of said cells and subsets into the circulating blood which is widely used to significantly increase in the blood the yield of circulating cells capable of regenerating hematopoiesis in vivo, thus minimizing the number of aphereses needed to achieve an engraftment dose.
- Other advantages of the methods and uses of the invention include the possibility of :

a) circumventing the need of general anasthesia,

b) harvesting even if iliac bones are damaged by previous radiotherapy or infiltrated with malignant cells,

c) achieving restoration of sustained hematopoietic functions more rapidly than with BM derived progenitor cells.

d) achieving restoration of sustained hematopoietic functions more rapidly and effectively than without a pre-treatment including a method or a use of the invention.

- Generally, methods and uses of the invention are effective and safe to mobilize to peripheral blood cells capable of regenerating hematopoiesis in vivo.

- Methods and uses of the invention are not toxic in view of main parameters of toxicity which are for example tumor growth, clinical and instrumental symptoms, or laboratory tests for cardiac, liver and renal function.

- The increased mobilization of circulating cells capable of regenerating hematopoiesis in vivo obtained with the methods and uses of the invention is extremely valuable in the context of hematopoietic stem cells transplantation by reducing the number of leukapheresis required to collect sufficient amount of hematopoietic cells to be transplanted.

- Methods and uses of the invention lead to a reduction of the volume of blood required to be processed during the apheresis or leukapheresis procedure in order to obtain the specified target number of cells. The advantages of processing a reduced volume of blood are that the patient spends less time on the cell separating machine, that it reduces the toxicity of the procedure, particularly in terms of the volume of anticoagulant to which the patient would be exposed during the procedure, that it reduces the machine and the operator's time.

- Furthermore, the transplantation of a population of blood cells enriched with cells capable of regenerating

hematopoiesis in vivo, which population is obtained from the peripheral blood by the methods or uses of the invention has the effect to enhance reconstitution of recipient's hematopoietic and immune systems following myeloablative or antiblastic therapies.

In a first aspect, the invention concerns a method of preparation of a population of circulating cells capable of regenerating hematopoiesis in vivo comprising :

a) administering to a donor a composition comprising growth hormone or one of its derivatives or any factor inducing growth hormone release in an amount sufficient to increase in said donor the number of circulating cells capable of regenerating hematopoiesis in vivo,
b) isolating a population of circulating cells capable of regenerating hematopoiesis in vivo from the peripheral blood of said donor.

The method of the invention thus produces a population of cells capable of regenerating hematopoiesis in vivo, this population being destined for transplantation in the same or in different individuals.

Thus, the invention concerns a method of preparation of a population of blood cells enriched with cells capable of regenerating hematopoiesis in vivo comprising :

a) administering to a donor a composition comprising growth hormone or one of its derivatives or any factor inducing growth hormone release in an amount sufficient to increase in said donor the number of circulating cells capable of regenerating hematopoiesis in vivo,
b) isolating a population of blood cells enriched with cells capable of regenerating hematopoiesis in vivo from the peripheral blood of said donor.

In a further embodiment, the invention relates to a method of isolating an increased number of circulating cells capable of regenerating hematopoiesis in vivo from a donor comprising :

a) administering to a donor a composition comprising growth hormone or one of its derivatives or any factor inducing growth hormone release alone, or in combination with other hematopoietic growth factors, to the subject in an amount sufficient to induce mobilization of cells capable of regenerating hematopoiesis in vivo to the peripheral blood,

b) isolating a population of blood cells enriched with cells capable of regenerating hematopoiesis in vivo from the peripheral blood of said donor.

In another embodiment, the invention concerns a method of preparation of a population of circulating cells capable of regenerating hematopoiesis in vivo comprising :

a) administering to a donor a composition comprising growth hormone or one of its derivatives or any factor inducing the growth hormone release in an amount sufficient to induce in said donor the mobilization or peripheralisation of circulating cells capable of regenerating hematopoiesis in vivo,

b) isolating a population of circulating cells capable of regenerating hematopoiesis in vivo from the peripheral blood of said donor or isolating a population of blood cells enriched with circulating cells capable of regenerating hematopoiesis in vivo from the peripheral blood of said donor.

Step b) [i.e. « isolating a population of (blood cells enriched with) circulating cells capable of regenerating hematopoiesis in vivo from the peripheral blood of said donor »] of the methods or uses of the invention may correspond to the operation of removing

peripheral blood from the donor wherein the number of cells capable of regenerating hematopoiesis in vivo has been increased by administration of growth hormone or one of its derivatives or any factor inducing growth hormone alone or in combination with other factors.

An amount sufficient to increase the number of circulating cells capable of regenerating hematopoiesis in vivo, an amount sufficient to induce the mobilization or peripheralisation of circulating cells capable of regenerating hematopoiesis in vivo or an amount sufficient to induce mobilization of cells capable of regenerating hematopoiesis in vivo to the peripheral blood can be administered in one or several doses during one or several days.

The operation of removing peripheral blood from the donor may correspond to leukapheresis. Leukapheresis is a procedure, in which, leukocytes are removed from the withdrawn blood and the remainder of the blood is retransfused into the donor.

Cells capable of regenerating hematopoiesis in vivo present in the isolated population of blood cells can be further purified in order to increase the concentration of said cells. Said purification may be done by positive selection of CD34 positive cells.

In a further embodiment, the invention concerns a method of preparation of a donor of circulating cells, which cells are capable of regenerating hematopoiesis in vivo comprising the administration to said donor of a composition comprising growth hormone or one of its derivatives or any factor inducing growth hormone release in an amount sufficient to increase the number of circulating cells capable of regenerating hematopoiesis in vivo in said donor.

In another embodiment, the invention concerns a method for increasing the number of circulating cells

capable of regenerating hematopoiesis in vivo in a donor by administration to said donor of a composition comprising growth hormone or one of its derivatives or any factor inducing the growth hormone release.

The term "increased" or "increase" and the term "enriched" generally mean in the context of the invention that the "increased" or "enriched" parameter (number) has a value which is above the standard value of this parameter. The standard value of the parameter is measured in a body or in a sample of a body which has not received any mobilising agent of cells capable of regenerating hematopoiesis in vivo. The standard value of the number of $CD34^+$ cells per microliter of blood is for example 3.8 (+ or - 3.2) cells per microliter of peripheral blood (Anderlini et al., 1997).

The circulating cells capable of regenerating hematopoiesis in vivo may be $CD34^+$ cells.

The frequency of $CD34^+$ cells in the blood may be measured by FACScan measurements (Siena et al., 1989 & 1991).

The increased number of $CD34^+$ cells in the peripheral blood of the donor or the level of enrichment of $CD34^+$ cells in the isolated preparation of blood cells may be more than 10, 25, 34 or 80 $CD34^+$ cells per microliter of peripheral blood.

The increased number of $CD34^+$ cells in the peripheral blood of the donor or the level of enrichment of $CD34^+$ cells in the isolated preparation of blood cells may be at least $2x10^6$ $CD34^+$ cells per kilogram of recipient body weight, or at least $4x10^6$ $CD34^+$ cells per kilogram of recipient body weight or at least $8x10^6$ $CD34^+$ cells per kilogram of recipient body weight.

The increased number of $CD34^+$ cells in the peripheral blood of the donor or the level of enrichment of $CD34^+$ cells in the isolated preparation of blood cells

may be at least $2x10^6$, $4x10^6$, $5x10^6$, $6x10^6$, $8x10^6$ or $15x10^6$ $CD34^+$ cells per kilogram of donor body weight.

There is a correlation between the number of $CD34^+$ cells required for transplantation and the corresponding GM-CFC activity which can be measured (Weaver et al., 1998). Therefore, the increased number of circulating cells capable of regenerating hematopoiesis in vivo or the level of enrichment of cells capable of regenerating hematopoiesis in vivo in the isolated preparation of blood cells may correspond to at least $1x10^5$ GM-CFC per kilogram of donor or recipient body weight.

The number of $CD34^+$ cells in the blood correlates well with CFU-GM (Siena et al., 1991). CFU-GM is the colony forming unit, granulocyte and macrophage. Therefore, the increased number of circulating cells capable of regenerating hematopoiesis in vivo or the level of enrichment of cells capable of regenerating hematopoiesis in vivo in the isolated preparation of blood cells may correspond to at least 500 CFU-GM per milliliter of peripheral blood.

With the same reasoning, the increased number of circulating cells capable of regenerating hematopoiesis in vivo or the level of enrichment of cells capable of regenerating hematopoiesis in vivo in the isolated preparation of blood cells may correspond to an increased level of CFU-C, CFU-Meg or BFU-E. CFU-C is the colony forming unit, culture ; CFU-Meg is the colony forming unit, megakaryocyte and BFU-E is the burst forming unit, erythroid.

The number of $CD34^+$ cells in the blood correlates well with the white blood cell count. Therefore, the increased number of circulating cells capable of regenerating hematopoiesis in vivo or the level of enrichment of cells capable of regenerating hematopoiesis in vivo in the isolated preparation of blood cells may

correspond to at least 1000 white blood cells per microliter of peripheral blood.

The circulating cells capable of regenerating hematopoiesis in vivo may be $CD34^+/CD33^+$ cells and/or $CD34^+/CD38^-$ cells and/or $CD34^+$/Thy-I cells and/or $CD34^+$/Thy-I/$CD38^-$ cells and/or $CD33^+$ cells and/or bone-marrow stem cells and/or progenitor cells and/or long-term culture initiating cells (LTC-IC) and/or cells that fulfill self renewal potential and/or cells that fulfill pluripotential characteristics and/or cells that initiate long term bone marrow culture and/or cells that can generate multiple cell lineages. Cell lineages may be fully differentiated blood cells.

The $CD34^+/CD38^-$ cells and $CD34^+$/Thy-I cells and $CD34^+$/Thy-I/$CD38^-$ cells are recited for example in *Anderlini et al* (see references). The $CD34^+/CD33^+$ cells and the $CD33^+$ cells are recited for example in *Siena et al* ; 1991 (see references). The long-term culture initiating cells (LTC-IC) are recited for example in *Heather et al*(see references). Cells that fulfill self renewal potential and/or cells that fulfill pluripotential characteristics and/or cells that initiate long term bone marrow culture are recited for example in *Anderlini et al*(see references).

In a further embodiment, the invention relates to the following uses :

- Use of growth hormone, one of its derivatives, or any factor inducing growth hormone release for increasing or expanding the number of circulating cells capable of regenerating hematopoiesis in vivo.
- Use of growth hormone, one of its derivatives or any factor inducing growth hormone release for peripheralizing cells capable of regenerating hematopoiesis in vivo.

- Use of growth hormone, one of its derivatives, or any factor inducing growth hormone release to prepare a medicament or a composition for increasing or expanding the number of circulating cells capable of regenerating hematopoiesis in vivo.
- Use of growth hormone, one of its derivatives or any factor inducing growth hormone release to prepare a medicament or a composition for peripheralizing cells capable of regenerating hematopoiesis in vivo.
- Use according to any one of the preceding uses wherein the circulating cells capable of regenerating hematopoiesis in vivo are $CD34^+$ cells.
- Use according to the preceding use wherein the increased number of $CD34^+$ cells is more than 10, 25, 34 or 80 $CD34^+$ cells per microliter of peripheral blood.
- Use according to any one of the preceding uses wherein the increased number of $CD34^+$ cells is at least $2x10^6$ $CD34^+$ cells per kilogram of recipient body weight, or at least $4x10^6$ $CD34^+$ cells per kilogram of recipient body weight or at least $8x10^6$ $CD34^+$ cells per kilogram of recipient body weight.
- Use according to any one of the preceding uses wherein the increased number of $CD34^+$ cells is at least $2x10^6$, $4x10^6$, $5x10^6$, $6x10^6$, $8x10^6$ or $15x10^6$ $CD34^+$ cells per kilogram of donor body weight.
- Use according to any one of the preceding uses wherein the increased number of circulating cells capable of regenerating hematopoiesis in vivo corresponds to at least $1x10^5$ GM-CFC per kilogram of donor or recipient body weight.
- Use according to any one of the preceding uses wherein the increased number of circulating cells capable of regenerating hematopoiesis in vivo corresponds to at least 500 CFU-GM per milliliter of peripheral blood.

- Use according to any one of the preceding uses wherein the increased number of circulating cells capable of regenerating hematopoiesis in vivo corresponds to an increased level of CFU-C, CFU-Meg or BFU-E.
- Use according to any one of the preceding uses wherein the increased number of circulating cells capable of regenerating hematopoiesis in vivo substantially corresponds to a white blood cell count which is at least 1000 cells per microliter of peripheral blood.
- Use according to any one of the preceding uses wherein the circulating cells capable of regenerating hematopoiesis in vivo are $CD34^+/CD33^+$ cells and/or $CD34^+/CD38^-$ cells and/or $CD34^+$/Thy-I cells and/or $CD34^+$/Thy-I/$CD38^-$ cells and/or $CD33^+$ cells and/or stem cells and/or progenitor cells and/or long-term culture initiating cells (LTC-IC) and/or cells that fulfill self renewal potential and/or cells that fulfill pluripotential characteristics and/or cells that initiate long term bone marrow culture.
- Use according to any one of the preceding uses wherein the medicament or composition comprises further one or several compound(s) chosen among the following groups of compounds : hematopoietic growth factors, cytokines, chemokines, monoclonal antibodies.
- Use according to any one of the preceding uses wherein the cytokines group comprises IL-1, IL-3, G-CSF, GM-CSF or SCF ; the chemokines group comprises MIP-1α or thrombopoietin (TPO) ; the monoclonal antibodies group comprises anti-VLA-4 antibodies.
- Use according to any one of the preceding uses wherein the medicament or composition comprises Growth Hormone and G-CSF.
- Use according to any one of the preceding uses wherein GH and G-CSF are administered separately and/or simultaneously.

- Use according to any one of the preceding uses wherein Growth Hormone is administered in an amount of around 33μg per kilogram of body weight.
- Use according to any one of the two preceding uses wherein the G-CSF is administered in an amount of around 5μg or around 10μg per kilogram.
- Use according to any one of the two preceding uses wherein the administration is made by intravenous or subcutaneous route.
- Use according to any one of the preceding uses wherein the administration is made by parenteral, subcutaneous, intravenous, intramuscular, intraperitoneal, transdermal or buccal routes.
- Use according to any one of the preceding uses wherein the administration is daily or three times a day.
- Use according to any one of the preceding uses wherein the administration of growth hormone is done three times a day and the administration of G-CSF is done daily.
- Use according to any one of the preceding uses wherein the administration is made over a period of 5 days or over a period of 10 days, until leukapheresis or until full recovery.
- Use according to any one of the preceding uses wherein the administration is made until leukapheresis or until full recovery.
- Use according to any one of the preceding uses wherein growth hormone is recombinant growth hormone.
- Use according to any one of the preceding uses wherein growth hormone is human growth hormone.

In a second aspect, the invention concerns a method of preparation of a population of circulating cells capable of regenerating hematopoiesis in vivo comprising :

a) administering to a donor a composition comprising growth hormone or one of its derivatives or any factor inducing growth hormone release in an amount sufficient to reduce the volume of blood required to be processed in order to obtain the specified target number of circulating cells capable of regenerating hematopoiesis in vivo,

b) processing or isolating said reduced volume of blood ; and optionally

c) isolating a population of circulating cells capable of regenerating hematopoiesis in vivo from said isolated volume.

Step b) or c) [isolating a population of circulating cells capable of regenerating hematopoiesis in vivo from said isolated volume] of the methods or uses of the invention may correspond to the operation of removing peripheral blood from the donor wherein the number of cells capable of regenerating hematopoiesis in vivo has been increased by administration of growth hormone or one of its derivatives or any factor inducing growth hormone alone or in combination with other factors.

An amount sufficient to reduce the volume of blood required to be processed in order to obtain the specified target number of circulating cells capable of regenerating hematopoiesis in vivo can be administered in one or several doses during one or several days.

The operation of removing peripheral blood from the donor may correspond to leukapheresis. Leukapheresis is a procedure, in which, leukocytes are removed from the withdrawn blood and the remainder of the blood is retransfused into the donor.

Cells capable of regenerating hematopoiesis in vivo present in the isolated population of blood cells can be further purified in order to increase the concentration

of said cells. Said purification may be done by positive selection of CD34 positive cells.

In another embodiment, the invention concerns a method of preparation of a donor of circulating cells, which cells are capable of regenerating hematopoiesis in vivo comprising the administration to said donor of a composition comprising growth hormone or one of its derivatives or any factor inducing growth hormone release in an amount sufficient to reduce the volume of blood required to be processed in order to obtain the specified target number of circulating cells capable of regenerating hematopoiesis in vivo and/or to reduce the number of leukapheresis required to collect sufficient amount of circulating cells capable of regenerating hematopoiesis in vivo to be tranplanted.

An amount sufficient to reduce the volume of blood required to be processed in order to obtain the specified target number of circulating cells capable of regenerating hematopoiesis in vivo and/or to reduce the number of leukapheresis required to collect sufficient amount of circulating cells capable of regenerating hematopoiesis in vivo to be tranplanted can be administered in one or several doses during one or several days.

The volume of blood required to be processed may be the volume of blood required to be processed during the apheresis or leukapheresis procedure.

In a further embodiment, the invention concerns a method for reducing the volume of blood required to be processed in order to obtain the specified target number of circulating cells capable of regenerating hematopoiesis in vivo in a donor and/or for reducing the number of leukapheresis required to collect sufficient amount of circulating cells capable of regenerating hematopoiesis in vivo to be tranplanted by administration

of a composition comprising the growth hormone or one of its derivatives or any factor inducing the growth hormone release to said donor .

The term "reduced" generally means in view of the invention that the "reduced" parameter (volume) has a value which is inferior to the standard value of this parameter.

The specified target number of circulating cells capable of regenerating hematopoiesis in vivo is at least $2x10^4$ LTC-IC per kg of donor or recipient body, around or more than $2x10^6$ $CD34^+$ cells per kilogram of donor or recipient body weight, around or more than $4x10^6$ $CD34^+$ cells per kilogram of donor or recipient body weight or around or more than $8x10^6$ $CD34^+$ cells per kilogram of donor or recipient body weight.

The required volume of blood may be comprised in a range of about 30 to about 900 milliliters.

In another embodiment, the invention relates to the following uses :

- Use of growth hormone, one of its derivatives or any factor inducing growth hormone release for reducing the volume of blood required to be processed in order to obtain the specified target number of circulating cells capable of regenerating hematopoiesis in vivo.
- Use of growth hormone, one of its derivatives or any factor inducing growth hormone release to prepare a medicament or a composition for reducing the volume of blood required to be processed in order to obtain the specified target number of circulating cells capable of regenerating hematopoiesis in vivo.
- Use according to the preceding use wherein the specified target number of circulating cells capable of regenerating hematopoiesis in vivo is around or more than $2x10^4$ LTC-IC per kg of donor or recipient body, around or more than $2x10^6$ $CD34^+$ cells per

kilogram of donor or recipient body weight, around or more than $4x10^6$ $CD34^+$ cells per kilogram of donor or recipient body weight or around or more than $8x10^6$ $CD34^+$ cells per kilogram of donor or recipient body weight.

- Use according to any one of the two preceding uses wherein the required volume of blood is comprised in a range of about 30 to about 900 milliliters.
- Use according to any one of the preceding uses wherein the medicament or composition comprises further one or several compound chosen among the following groups of compounds : hematopoietic growth factors, cytokines, chemokines, monoclonal antibodies.
- Use according to any one of the preceding uses wherein the cytokines group comprises IL-1, IL-3, G-CSF, GM-CSF or SCF ; the chemokines group comprises MIP-1α or thrombopoietin (TPO) ; the monoclonal antibodies group comprises anti-VLA-4 antibodies.
- Use according to any one of the preceding uses wherein the medicament or composition comprises Growth Hormone and G-CSF.
- Use according to any one of the preceding claims wherein GH and G-CSF are administered separately and/or simultaneously.
- Use according to any one of the preceding uses wherein Growth Hormone is administered in an amount of around 33 μg per kilogram of body weight.
- Use according to any one of the two preceding uses wherein the G-CSF is administrated in an amount of around 5 μg or around 10μg per kilogram.
- Use according to any one of the two preceding uses wherein the administration is made by intravenous or subcutaneous route.
- Use according to any one of the preceding uses wherein the administration is made by parenteral,

subcutaneous, intravenous, intramuscular, intraperitoneal, transdermal or buccal routes.

- Use according to any one of the preceding uses wherein the administration is daily or three times a day.
- Use according to any one of the preceding claims wherein the administration of growth hormone is done three times a day and the administration of G-CSF is done daily.
- Use according to any one of the preceding uses wherein the administration is made over a period of 5 days or over a period of 10 days, until leukapheresis or until full recovery.
- Use according to any one of the preceding claims wherein the administration is made until leukapheresis or until full recovery.
- Use according to any one of the preceding uses wherein the administration(s) is(are) made after chemotherapy, radiotherapy, myelosuppressive therapy, transplantation of cells capable of regenerating hematopoiesis in vivo or transplantation of bone-marrow.
- Use according to any one of the preceding uses wherein the administration(s) begin(s) around 7 days after the beginning of the chemotherapeutic treatment or around 2 days after the end of the chemotherapeutic treatment.
- Use according to any one of the preceding uses wherein the growth hormone is recombinant growth hormone.
- Use according to any one of the preceding uses wherein the growth hormone is human growth hormone.

In this application :

- The term "circulating" may be replaced by the term "blood" or "peripheral blood".

- The term "preparation" in the expression "method of preparation" may be replaced by "pre-treament" or by "preparation for blood extraction or leukapheresis".
- A "donor" as recited in the methods or uses of the invention may be a human or an animal, a healthy or a sick individual (patient). Said animal is preferably a mammal and may be chosen from domestic animals such as dogs, cats etc. or animals such as horses, cattle, sheep.
- The term « hematopoiesis » can mean the formation of the blood cells.
- The term "Growth hormone" encompasses human growth hormone (hGH) and all the homologous proteins of human growth hormone of different species and all the homologs of human growth hormone in species other than human. Species other than human may be any sort of domestic animal or horse for example.

In a preferred embodiment, growth hormone is human growth hormone. Human growth hormone (hGH), also known as somatotropin is a protein hormone produced and secreted by the somatotropic cells of the anterior pituitary. hGH plays a key role in somatic growth through its effects on the metabolism of proteins, carbohydrates and lipids.

Human growth hormone is a single polypeptide chain of 101 amino acids having two disulfide bonds, one between Cys-53 and Cys-165, forming a large loop in the molecule, and the other between Cys-182 and Cys-189, forming a small loop near the C-terminus.

The term « derivative » in the expression « derivatives of growth hormone » signifies in the context of the invention, molecules which differ structurally from GH but which conserve the function of GH with respect to its direct or indirect effect on the metabolism of proteins, carbohydrates and lipids and/or its mobilisation effect and/or recovery effect (i.e.

« mobilization or peripheralisation of circulating cells capable of regenerating hematopoiesis in vivo, increase of the number of circulating cells capable of regenerating hematopoiesis in vivo, reduction of the number of leukapheresis required to collect sufficient amount of circulating cells for transplantation, reduction of the volume of blood required to be processed in order to obtain the specified target number of circulating cells capable of regenerating hematopoiesis in vivo »)

Derivatives of human growth hormone (hGH) included in the invention include naturally-occurring derivatives, variants and metabolic products, degradation products primarily of biosynthetic hGH and engineered derivatives of hGH produced by genetic methods. Any derivative of hGH can be used for the purpose of the present invention as long as it retains the biological activity of hGH in view of the invention.

Examples of derivatives are splice variants, oligomers, aggregates, proteolytic cleavage products, variants having substitutions, insertions or deletions of one or more amino acids etc.

Methionyl hGH is an example of derivative of hGH which is produced through recombinant DNA technology. This compound is actually a derivative of hGH having one additional methionine residue at its N-terminus (Goeddel et al., 1979).

Another example of derivative of hGH is a naturally occurring variant of hGH called 20-K-hGH which has been reported to occur in the pituitary as well as in the bloodstream (Lewis et al, 1978 ; Lewis et al, 1980). This compound, which lacks the 15 amino acid residues from Glu-32 to Gln-46, arises from an alternative splicing of the messenger ribonucleic acid (DeNoto et al., 1981).

Another example of derivative of hGH is acetylated at the N-terminus (Lewis et al., 1979).

Human growth hormone may further be in a monomeric, dimeric and higher molecular weight oligomeric form or in a mixture of said forms.

Human growth hormone may be in aggregated forms found both in the pituitary and in the circulation (Stolar et al., 1984 ; Stolar and Baumann, 1986).

The dimeric form of hGH may be of distinct types :

- a disulfide dimer connected through interchain disulfide bonds (Lewis et al., 1977),
- a covalent or irreversible dimer that is detected on sodium dodecylsulfate-polyacrylamide gels and that is not a disulfide dimer (Bewley and Li, 1975), and
- a non-covalent dimer which is easily dissociated into monomeric hGH by treatment with agents that disrupt hydrophobic interactions in proteins (Becker et al., 1987),
- a dimeric complex with Zn^{2+} (Cunningham et al., 1991).

Scatchard analysis has revealed that two Zn^{2+} ions associate per hGH dimer in a cooperative fashion, and this Zn^{2+}-hGH dimeric complex was found to be more stable to denaturation than monomeric hGH (Cunningham et al., 1991).

A number of derivatives of hGH arise from proteolytic modifications of the molecule. The primary pathway for the metabolism of hGH involves proteolysis. The region of hGH around residues 130-150 is extremely susceptible to proteolysis, and several derivatives of hGH having nicks or deletions in this region have been described (Thorlacius-Ussing, 1987). This region is in the large loop of hGH, and cleavage of a peptide bond there results in the generation of two chains that are connected through the disulfide bond at Cys-53 and Cys-

165. Many of these two-chain forms are reported to have increased biological activity (Singh et al., 1974).

Many derivatives of human growth hormone have been generated artificially through the use of enzymes. The enzymes trypsin and subtilisin, as well as others, have been used to modify hGH at various points throughout the molecule (Lewis et al., 1977). One such derivative, called two-chain anabolic protein (2-CAP), was formed through the controlled proteolysis of hGH using trypsin.

Another example of derivative of hGH is deamidated hGH. Asparagine and glutamine residues in proteins are susceptible to deamidation reactions under appropriate conditions. An example of deamidated hGH is pituitary hGH which has been shown to undergo this type of reaction, resulting in conversion of Asn-152 to aspartic acid and also, to a lesser extent, conversion of Gln-137 to glutamic acid (Lewis et al., 1981). Another example of deamidated hGH is Biosynthetic hGH which is known to degrade under certain storage conditions, resulting in deamidation at a different asparagine (Asn-149). This is the primary site of deamidation, but deamidation at Asn-152 is also seen (Becker et al., 1988). Deamidation at Gln-137 has not been reported in biosynthetic hGH.

Another example of derivative of hGH is sulfoxide hGH. Methionine residues in proteins are susceptible to oxidation, primarily to the sulfoxide. Both pituitary-derived and biosynthetic hGH undergo sulfoxidations at Met-14 and Met-125 (Becker et al., 1988). Oxidation at Met-170 has also been reported in pituitary but not biosynthetic hGH.

Another example of derivative of hGH is truncated forms of hGH which have been produced, either through the actions of enzymes or by genetic methods. 2-CAP, generated by the controlled actions of trypsin, has the first eight residues at the N-terminus of hGH removed.

Other truncated versions of hGH have been produced by modifying the gene prior to expression in a suitable host. The first 13 residues have been removed to yield a derivative having distinctive biological properties in which the polypeptide chain is not cleaved (Gertler et al., 1986).

hGH and its derivatives may be produced by recombinant DNA technology which permits production of an unlimited supply of hGH in a number of different systems. Purification of hGH or its derivatives from the culture medium is facilitated by low amounts of contaminating proteins present. In fact, it has been shown that hGH can be purified on a laboratory scale by a single purification step on a reversed-phase HPLC column .

Recombinant hGH is generally marketed as vials containing hGH plus additional excipients, e.g., glycine and mannitol, in a lyophilized form. A companion diluent vial is provided, allowing the patient to reconstitute the product to the desired concentration prior to administration of the dose.

In general, no significant differences have been observed in the pharmacokinetics or biological activities of recombinant natural sequence hGH, recombinant N-methionyl-hGH, or pituitary-derived material in humans (Moore et al., 1988 ; Jorgensen et al. , 1988).

The human growth hormone as used in the present invention can include functional derivatives as noted above, as well as other types of derivatives, fragments, variants, analogs, or chemical derivatives. A functional derivative retains at least a portion of the amino acid sequence of hGH which permits its utility in accordance with the present invention, namely mobilization of circulating cells capable of regenerating hematopoiesis in vivo for example.

In the meaning of the invention, a « derivative » may be :

- A "fragment" of the human growth hormone according to the present invention refers to any subset of the molecule, that is, a shorter peptide.
- A "variant" of the human growth hormone according to the present invention refers to a molecule which is substantially similar to either the entire peptide or a fragment thereof. Variant peptides may be conveniently prepared by direct chemical synthesis of the variant peptide, using methods well known in the art.

Alternatively, amino acid variants of hGH can be prepared by mutations in the cDNA encoding the synthetized hGH derivatives. Such variants comprise deletions, insertions or substitution of residues within the amino acid sequence. Any combination of deletions, insertions, and subtitutions may also be made, provided that the final construct possesses the desired activity.

At the genetic level, these variants ordinarily are prepared by site-directed mutageneis (as exemplified by (Adelman et al., 1983)) of nucleotides in the DNA encoding the peptide molecule, thereby producing DNA encoding the variant, and thereafter expressing the DNA in recombinant cell culture. The variants typically exhibit the same biological activity as the non-variant peptide.

- An "analog" of human growth hormone according to the present invention refers to a non-natural molecule which is substantially similar to either the entire molecule or to an active fragment thereof.
- A "chemical derivative" of human growth hormone according to the present invention contains additional chemical moieties not normally part of the human growth hormone derivative amino acid sequence. Covalent modifications of the amino acid sequence are included

within the scope of this invention. Such modifications may be intrcduced into the human growth hormone by reacting targeted amino acid residues of the peptide with an organic derivatizing agent that is capable of reacting with selected side chains or terminal residues.

The types of substitutions which may be made in the human growth hormone according to the present invention may be based on analysis of the frequencies of amino acid changes between a homologous protein of different species. Based upon such analysis, conservative substitutions may be defined herein as exchanges within one of the following five groups :

I : Small, aliphatic, nonpolar or slightly polar residues : Ala, Ser, Thr, Pro, Gly

II : Polar, negatively-charged residues and their amides : Asp, Asn, Glu, Gln

III : Polar, positively-charged residues : His, Arg, Lys

IV : Large, aliphatic non-polar residues : Met, Leu, Ile, Val, Cys

V : Large aromatic residues : Phe, Try, Trp

Within the foregoing groups, the following substitutions are considered to be "highly conservative" :

- Asp/Glu
- His/Arg/Lys
- Phe/Tyr/Trp
- Met/Leu/Ile/Val

Semi-conservative substitutions are defined to be exchanges between two of groups (I)-(IV) above which are limited to supergroup (A), comprising (I), (II), and (III) above, or to supergroup (B), comprising (IV) and (V) above. Substitutions are not limited to the genetically encoded or even the naturally- occurring amino acids. When the epitope is prepared by peptide synthesis, the desired amino acid may be used directly.

Alternatively, a genetically encoded amino acid may be modified by reacting it with an organic derivatizing agent that is capable of reacting with selected side chains or terminal residues.

Cysteinyl residues most commonly are reacted with alpha- haloacetates (and corresponding amines), such as chloroacetic acid or chloroacetamide, to give carboxylmethyl or carboxyamidomethyl derivatives. Cysteinyl residues also are derivatized by reaction with bromotrifluoroacetone, alpha-bromo-beta-(5-imidazoyl)propionic acid, chloroacetyl phosphate, N-alkylmaleimides, 3-nitro-2-pyridyl disulfide, methyl-2-pyridyl disulfide, p-chloromercuribenzoate, 2-chloromercuri-4-nitrophenol, or chloro-7-nitrobenzo-2-oxa-1,3-diazole.

Histidyl residues are derivatized by reaction with diethylprocarbonate at pH 5.5-7.0 because this agent is relatively specific for the histidyl side chain. Parabromophenacyl bromide is also useful ; the reaction is preferably performed in 0.1 M sodium cacodylate at pH 6.0.

Lysinyl and amino terminal residues are reacted with succinic or other carboxylic acid anhydrides. Derivatization with these agents has the effect of reversing the charge of the lysinyl residues. Other suitable reagents for derivatizing alpha-amino acid-containing residues include imidoesters such as methyl picolinimidate ; pyridoxal phosphate ; pyridoxal ; chloroborohydride; trinitrobenzenesulfonic acid ; O-methyliosurea ; 2,4-pentanedione ; and transaminase-catalyzed reaction with glyoxylate.

Arginyl residues are modified by reaction with one or several conventional reagents, among them phenylglyoxal ; 2,3- butanedione ; and ninhydrin. Derivatization of arginine residues requires that the

reaction be performed in alkaline conditions because of the high pKa of the guanidine functional group. Furthermore, these reagents may react with the groups of lysine, as well as the arginine epsilon-amino group.

The specific modification of tyrosyl residues per se has been studied extensively, with particular interest in introducing spectral labels into tyrosyl residues by reaction with aromatic diazonium compounds or tetranitromethane. Most commonly, N-acetylimidazole and tetranitromethane are used to form O-acetyl tyrosyl species and e-nitro derivatives, respectively.

Carboxyl side groups (aspartyl or glutamyl) are selectively modified by reaction with carbodiimides (R'N-C-N-R') such as 1-cyclohexyl-3-[2-morpholinyl-(4-ethyl)]carbodiimide or 1-ethyl-3-(4-azonia-4,4-dimethylpentyl)carbodiimide. Furthermore, aspartyl and glutamyl residues are converted to asparaginyl and glutaminyl residues by reaction with ammonium ions.

Glutaminyl and asparaginyl residues are frequently deamidated to the corresponding glutamyl and aspartyl residues. Alternatively, these residues are deamidated under mildly acidic conditions. Either form of these residues falls within the scope of this invention.

While the present invention may be carried out with recombinant human growth hormone derivatives made by recombinant DNA technology, for instance in procaryotic or eucaryotic cells, these derivatives can also be made by conventional protein synthesis methods which are well known to those skilled in the art.

Growth hormone may be a protein, a peptide, a DNA molecule, a RNA molecule. DNA molecule and RNA molecule may encode hGH and all its derivatives including those recited above.

Growth hormone may preferably be recombinant growth hormone.

Determination of amounts of growth hormone, of one of its derivatives or of any factor inducing growth hormone release to be administered in a method or use of the invention described above is within the skill of the art.

Typical dosage of growth hormone, of one of its derivatives or of any factor inducing growth hormone release will start at about 1 microgram per kilogram of the patient weight per day and dose will be escalated until the desired effect (mobilization or peripheralisation of circulating cells capable of regenerating hematopoiesis in vivo, increase of the number of circulating cells capable of regenerating hematopoiesis in vivo, reduction of the number of leukapheresis required to collect sufficient amount of circulating cells for transplantation, reduction of the volume of blood required to be processed in order to obtain the specified target number of circulating cells capable of regenerating hematopoiesis in vivo) is reached.

The dosage of growth hormone, of one of its derivatives or of any factor inducing growth hormone release administered depends upon the age, sex, health and weight of the donor, type of previous or concurrent treatment, if any, frequency of the treatment and the nature of the effect desired.

Growth hormone or one of its derivatives may advantageously be administered in an amount comprised between 20 and 50 μg per kilogram of body weight, more particularly between 30 and 40 μg per kilogram of body weight.

A preferred dosage of Growth hormone or one of its derivatives to be administered is around 33 μg per kilogram of body weight.

Growth hormone or its derivatives or any factor inducing growth hormone release may be administered alone or in conjunction or association with other factors.

Growth hormone or its derivatives or any factor inducing growth hormone release may advantageously be present in a composition which comprises further one or several compound(s) chosen among the compounds belonging to the following groups : hematopoietic growth factors, cytokines, chemokines, monoclonal antibodies.

Growth hormone or its derivatives or any factor inducing growth hormone release and one or several compound(s) chosen among the compounds belonging to the following groups : hematopoietic growth factors, cytokines, chemokines, monoclonal antibodies can be administered simultaneously or at different times and/or at the same site or at different site(s) and/or in the same or in a different composition or medicament.

The cytokine group can comprise IL-1,IL-3, G-CSF, GM-CSF or SCF. The chemokine group can comprise MIP-1α or thrombopoietin (TPO). The monoclonal antibody group can comprise anti-VLA-4 antibodies.

Preferably, Growth hormone or its derivatives or any factor inducing growth hormone is present in a composition which comprises granulocyte colony stimulating factor (G-CSF).

Preferably, Growth hormone or its derivatives or any factor inducing growth hormone is associated with G-CSF.

Growth hormone or its derivatives or any factor inducing growth hormone and G-CSF can be administered simultaneously or at different times and/or at the same site or at different site(s) and/or in the same or in a different composition or medicament.

Growth hormone or its derivatives or any factor inducing growth hormone release and G-CSF may advantageously be administered separately.

G-CSF may advantageously be administered in an amount comprised between 3 and 15 μg per kilogram of body weight, more particularly between 4 and 12 μg per kilogram of body weight.

A preferred dosage of G-CSF to be administered is around 5μg or around 10μg per kilogram of body weight.

In a preferred embodiment, Growth hormone or one of its derivatives is administered in an amount comprised between 20 and 50 μg per kilogram of body weight, more particularly between 30 and 40 μg per kilogram of body weight and G-CSF is administered in an amount comprised between 3 and 15 μg per kilogram of body weight, more particularly between 4 and 12 μg per kilogram of body weight.

In a preferred embodiment, Growth hormone or one of its derivatives is administered in an amount of 33 μg per kilogram of body weight and G-CSF is administered in an amount of around 5μg or around 10μg per kilogram of body weight.

According to the invention, the expression « administration in an amount sufficient to increase the number of circulating cells capable of regenerating hematopoiesis in vivo or to reduce the volume of blood required to be processed in order to obtain the specified target number of circulating cells capable of regenerating hematopoiesis in vivo » can mean one or several administration(s), one or several times a day and during one or several days for a cumulated amount sufficient to increase the number of circulating cells capable of regenerating hematopoiesis in vivo or to reduce the volume of blood required to be processed in order to obtain the specified target number of

circulating cells capable of regenerating hematopoiesis in vivo.

The pharmaceutical compositions or compositions which are used in the methods and uses of the invention are in a pharmaceutical acceptable form optionally combined with an acceptable carrier.

These compositions can be administered by any means that achieve their intended purposes.

The compositions used in the methods or uses of the invention may be administered alone or in conjunction with other therapeutics directed to a disease or directed to other symptoms thereof.

The compositions used in the methods or uses of the invention may be administered by the intravenous or the subcutaneous route.

After intravenous administration, the elimination of hGH is described by first-order kinetics with a serum half-life of 12- 30 minutes in both animals and humans (Moore et al., 1988; Hendricks et al., 1985). Traditionally, intramuscular injection has been the method of choice as the preferred route of delivery. In humans, absorption of exogenous hGH appears to be more rapid from the intramuscular site, with a time to maximum concentration of two to three hours, compared to four to six hours after subcutaneous administration. The disappearance phase from serum has been reported to range from 12-20 hours for intramuscular administration, and 20-24 hours after subcutaneous administration (Albertsson-Wikland et al., 1986; Jorgensen et al., 1987).

The compositions used in the methods or uses of the invention may be administered by parenteral routes such as subcutaneous, intravenous, intramuscular, intraperitoneal, or transdermal route or by mucosal routes such as buccal or oral route.

The composition comprising growth hormone or one of its derivatives or any factor inducing growth hormone release may be administered by parenteral routes such as subcutaneous, intravenous, intramuscular, intraperitoneal, or transdermal route or by mucosal routes such as buccal or oral route.

Preferably, growth hormone or one of its derivatives or any factor inducing growth hormone release is administered subcutaneously.

The total dose or amount required for each treatment, method or use of the invention may be administered in multiple or single dose.

The composition comprising growth hormone or one of its derivatives or any factor inducing growth hormone release may be administered daily or three times a day.

Preferably, the composition comprising growth hormone or one of its derivatives or any factor inducing growth hormone release is administered three times a day.

Preferably, growth hormone or one of its derivatives or any factor inducing growth hormone release is administered daily or three times a day.

In a preferred embodiment, growth hormone or one of its derivatives or any factor inducing growth hormone release is administered three times a day.

If a method or use of the invention comprises the administration of growth hormone or one of its derivatives or any factor inducing growth hormone release and G-CSF, G-CSF is preferably administered once a day and/or subcutaneously.

If a method or use of the invention comprises the administration of growth hormone or one of its derivatives or any factor inducing growth hormone release and G-CSF, growth hormone or one of its derivatives or any factor inducing growth hormone release is preferably

administered three times a day and G-CSF is preferably administered daily.

The composition comprising growth hormone or one of its derivatives or any factor inducing growth hormone release may be a daily administration that can start up to 20 days pre-leukapheresis.

The composition comprising growth hormone or one of its derivatives or any factor inducing growth hormone release may be administered over a period of 5 days or over a period of 10 days, until leukapheresis or until the desired effect (mobilization or peripheralisation of circulating cells capable of regenerating hematopoiesis in vivo, increase of the number of circulating cells capable of regenerating hematopoiesis in vivo, reduction of the number of leukapheresis required to collect sufficient amount of circulating cells for transplantation, reduction of the volume of blood required to be processed in order to obtain the specified target number of circulating cells capable of regenerating hematopoiesis in vivo) is reached.

Preferably, the composition comprising growth hormone or one of its derivatives or any factor inducing growth hormone release is administered until leukapheresis and/or the desired effect (mobilization or peripheralisation of circulating cells capable of regenerating hematopoiesis in vivo, increase of the number of circulating cells capable of regenerating hematopoiesis in vivo, reduction of the number of leukapheresis required to collect sufficient amount of circulating cells for transplantation, reduction of the volume of blood required to be processed in order to obtain the specified target number of circulating cells capable of regenerating hematopoiesis in vivo) is reached.

Methods and uses of the invention are advantageously carried out after chemotherapy, radiotherapy, myelosuppressive therapy, transplantation or engraftment of cells capable of regenerating hematopoiesis in vivo or transplantation of bone-marrow.

Methods and uses of the invention are advantageously carried out around 7 days after the beginning of a chemotherapeutic treatment or around 2 days after the end of a chemotherapeutic treatment.

In a preferred embodiment, growth hormone or one of its derivatives or any factor inducing growth hormone release and G-CSF are administered until leukapheresis, until mobilization or peripheralisation of circulating cells capable of regenerating hematopoiesis in vivo, until increase of the number of circulating cells capable of regenerating hematopoiesis in vivo, until reduction of the number of leukapheresis required to collect sufficient amount of circulating cells for transplantation, and/or until reduction of the volume of blood required to be processed in order to obtain the specified target number of circulating cells capable of regenerating hematopoiesis in vivo. In this preferred embodiment, growth hormone or one of its derivatives or any factor inducing growth hormone release is preferably administered three times a day and G-CSF is preferably administered once a day.

Methods and uses of the invention may be combined with a prior treatment called « chemopriming ». The « chemopriming » regimens which may be used are :

- high-dose cyclophosphamide (4 g/m^2) for patients with breast cancer or multiple myeloma,
- ifosfamide, etoposide for patients with non-Hodgkin's lymphoma or Hodgkin's disease,
- cyclophosphamide, etoposide, cisplatin (CVP) for patients with solid tumors (e.g., breast cancer).

To enhance the induction of cells capable of regenerating hematopoiesis in vivo rebound, methods and uses of the invention are started shortly after completion of chemopriming treatment and continued until completion of apheresis (5 to 12 μg/kg/d).

It is also noteworthy that in patients whose marrow stem cell pool is significantly diminished by prior chemotherapy, an additional chemopriming regimen might impair rather than induce cells capable of regenerating hematopoiesis in vivo peripheralization. Stem cells toxic chemotherapeutic agents such as busulfan, doxorubucin, melphalan, thiotepa and possibly fludarabine (and others) should not be part of a chemopriming regimen. On the other hand, cyclophosphamide is considered the ideal chemopriming drug with the least cells capable of regenerating hematopoiesis in vivo toxicity, although cardiotoxicity (dose > $4g/m^2$) and hemorrhagic cystitis are the well-known dose-limiting extramedullary side effects (Shepperd et al., 1990).

The population of blood cells enriched with cells capable of regenerating hematopoiesis in vivo obtained from the peripheral blood by the methods and uses of the invention can be re-infused, grafted or transplanted into the same individual which is in this case the donor (autologous transplantation) or into different individuals (nonautologous transplantation).

The population of blood cells enriched with cells capable of regenerating hematopoiesis in vivo obtained from the peripheral blood by the methods and uses of the invention are advantageously infused into an individual which has previously received one or several chemotherapy, radiotherapy, myelosuppressive, myeloablative or myelotoxic therapy.

Said operation of re-infusion, engraftment or transplantation belongs to the so-called Hematopoietic

Stem Cells Transplantation (HSCT) procedures. HSCT is a clinical procedure in which cells capable of regenerating hematopoiesis in vivo, obtained from bone marrow or peripheral blood, are transplanted to a patient.

An autologous transplantation is a transplantation in which donor and recipient are the same individual whereas a nonautologous transplantation is a transplantation in which donor and recipient are different individuals. The method of the invention encompasses both autologous and non-autologous transplantation.

In another part , the invention provides a method or a use of growth hormone or one of its derivatives or any factor inducing growth hormone release to enhance the mobilization or peripheralisation effect of G-CSF.

The invention provides a method or a use of growth hormone or one of its derivatives or any factor inducing growth hormone release to enhance mobilisation of circulating cells capable of regenerating hematopoiesis in vivo by G-CSF, to enhance increase the number of circulating cells capable of regenerating hematopoiesis in vivo by G-CSF, to enhance reduction of the number of leukapheresis required to collect sufficient amount of circulating cells for transplantation by G-CSF, and/or to enhance reduction of the volume of blood required to be processed in order to obtain the specified target number of circulating cells capable of regenerating hematopoiesis in vivo by G-CSF.

Thus, the administration of growth hormone or one of its derivatives or any factor inducing growth hormone release and G-CSF enhances or increases synergistically the mobilisation of circulating cells capable of regenerating hematopoiesis in vivo, enhances or increases synergistically the number of circulating cells capable of regenerating hematopoiesis in vivo, reduces the number

of leukapheresis required to collect sufficient amount of circulating cells for transplantation, and/or reduces the volume of blood required to be processed in order to obtain the specified target number of circulating cells capable of regenerating hematopoiesis in vivo with respect to the effect(s) obtained by administering G-CSF alone or without growth hormone or one of its derivatives or any factor inducing growth hormone release.

The administration of growth hormone or one of its derivatives or any factor inducing growth hormone release and G-CSF allows to use lower dose(s) of G-CSF than if G-CSF is used alone or without growth hormone or one of its derivatives or any factor inducing growth hormone release.

The administration of growth hormone or one of its derivatives or any factor inducing growth hormone release and G-CSF can be carried out simultaneously or at different times and/or at the same site or at different site(s) and/or in the same or in a different composition or medicament.

In a second part, the invention provides new uses for enhancing hematopoietic reconstitution. The invention relates to an agent capable of promoting, enhancing or accelerating the hematopoietic regeneration, recovery or reconstitution. The invention provides new uses for enhancing hematopoietic reconstitution.

Thus, the invention relates to the use of human growth hormone or one of its derivatives or any factor inducing human growth hormone release to prepare a medicament for enhancing hematopoietic reconstitution, in a human being.

Throughout the application, the term « enhancing » and all terms having the same root may be replaced by the term « promoting » or by the term « accelerating ».

Throughout the application, the term « reconstitution » and all terms having the same root may be replaced by the term « recovery » or by the term « regeneration ».

In another embodiment, the invention relates to the use of human growth hormone or one of its derivatives or any factor inducing human growth hormone release to prepare a medicament for enhancing hematopoietic reconstitution following bone marrow transplantation in a human being.

In another embodiment, the invention relates to the use of human growth hormone or one of its derivatives or any factor inducing human growth hormone release to prepare a medicament for enhancing engraftment of bone marrow or cells capable of regenerating hematopoiesis in vivo in a human being.

Another embodiment of the invention is the use of growth hormone or one of its derivatives or any factor inducing growth hormone release to prepare a medicament for enhancing hematopoietic reconstitution after transplantation of cells capable of regenerating hematopoiesis in vivo.

In a further embodiment, the invention relates to the use of growth hormone or one of its derivatives or any factor inducing growth hormone release to prepare a medicament for enhancing engraftment of cells capable of regenerating hematopoiesis in vivo.

Growth hormone may advantageously be human growth hormone.

Growth hormone and its derivatives may correspond to growth hormone and its derivatives which are recited above in this application in connection with the first part of the invention.

The hematopoietic reconstitution or the enhanced engraftment may be detected by an increase of the

peripheral White Blood Cell (WBC) count and/or granulocytes count and/or lymphocytes count and/or platelet count and/or erythrocyte count.

An increase of the peripheral White Blood Cell (WBC) count and/or granulocytes count and/or lymphocytes count and/or platelet count and/or erythrocyte count may be detected by comparison with the rate of increase of said counts in an individual who has received the same transplantation regimen but who has not received any hematopoietic reconstitution treatment.

The hematopoietic reconstitution or the enhanced engraftment may be detected by reduction of the period of time necessary to recover a normal or standard peripheral White Blood Cell (WBC) count and/or granulocytes count and/or neutrophil count and/or lymphocytes count and/or platelet count and/or erythrocyte count.

A normal or standard peripheral White Blood Cell (WBC) count and/or granulocytes count and/or neutrophil count and/or lymphocytes count and/or platelet count and/or erythrocyte count is the one which is measured in a healthy individual or in an individual which has not received any myeloablative, myelotoxic or myelosuppressive therapy, any chemotherapy, any radiotherapy or any transplantation.

A normal neutrophil count may be at least $0.5x10^9$ neutrophil cells per liter of peripheral blood.

A normal platelet count may be at least $20x10^9$ per liter of peripheral blood.

The hematopoietic reconstitution or enhancing engrafment may be detected by a reduction of the extent and/or duration of neutropenia and/or thrombocytopenia and/or anemia and/or hemorrhages and/or duration of prophylaxis.

The hematopoietic reconstitution or the enhancing engrafment may be detected by a reduction of the duration and/or severity of fever and/or infections.

A reduction of the extent and/or duration of neutropenia and/or thrombocytopenia and/or anemia and/or hemorrhages and/or duration of prophylaxis or a reduction of the duration and/or severity of fever and/or infections may be compared with said extent and/or duration and/or severity measured in an individual who has received the same transplantation regimen, the same chemotherapy, the same radiotherapy or the same myelosuppressive, myeloablative or myelotoxic therapy but who has not received any hematopoietic reconstitution treatment.

The hematopoietic reconstitution or the enhancing engrafment may be detected by a recovery of granulocytes which is at least 1000 per microliter of peripheral blood.

The hematopoietic reconstitution or the enhancing engrafment may be detected by a recovery of platelet count which is at least 50,000 per microliter of peripheral blood.

In another embodiment, the invention relates to the use of growth hormone or one of its derivatives or any factor inducing growth hormone release to prepare a medicament for treating a neoplastic disease, an hematological disorder, malignancies, Severe Combined Immune Deficiencies (SCIDs), congenitally or genetically determined hematopoietic abnormalities, anemia, aplastic anemia, leukemia and/or osteopetrosis.

A neoplastic disease may be breast cancer.

In another embodiment, the invention relates to the use of growth hormone or one of its derivatives or any factor inducing growth hormone release to prepare a medicament for reducing the bone marrow aplasia period

which follows transplantation, chemotherapy, radiotherapy or myeloablative, myelosuppressive or myelotoxic therapy, for preventing or treating opportunistic infections after transplantation, chemotherapy, radiotherapy or myeloablative, myelosuppressive or myelotoxic therapy or for limiting the risk of tumor recurrence after transplantation, chemotherapy, radiotherapy or myeloablative, myelosuppressive or myelotoxic therapy.

In a further embodiment, the invention relates to the use of growth hormone or one of its derivatives or any factor inducing growth hormone release to prepare a medicament for preventing or treating secondary effects of myeloablative, myelotoxic or myelosuppressive therapy and/or radiotherapy and/or chemotherapy and/or transplantation.

In a further embodiment, the invention relates to the use of growth hormone or one of its derivatives or any factor inducing growth hormone release to prepare a medicament for preventing or treating neutropenia and/or thrombocytopenia.

In a further embodiment, the invention relates to the use of growth hormone or one of its derivatives or any factor inducing growth hormone release to prepare a medicament for preventing or treating anemia following radiotherapy and/or chemotherapy and /or hematopoietic stem cells transplantation and/or transplantation of cells capable of regenerating hematopoiesis and/or Bone marrow transplantation and/or myelosuppressive or myelotoxic therapy.

In a further embodiment, the invention relates to the use of growth hormone or one of its derivatives or any factor inducing growth hormone release to prepare a medicament for preventing or treating neutropenia, following radiotherapy and/or chemotherapy and /or hematopoietic stem cells transplantation and/or

transplantation of cells capable of regenerating hematopoiesis and/or Bone marrow transplantation and/or myelosuppressive or myelotoxic therapy.

In a further embodiment, the invention relates to the use of growth hormone or one of its derivatives or any factor inducing growth hormone release to prepare a medicament for preventing or treating thrombocytopenia, following radiotherapy and/or chemotherapy and /or hematopoietic stem cells transplantation and/or transplantation of cells capable of regenerating hematopoiesis and/or Bone marrow transplantation and/or myelosuppressive or myelotoxic therapy.

The cells capable of regenerating hematopoiesis in vivo may belong to one or several of the following groups of cells : $CD34^+$ cells, $CD34^+CD33^+$ cells, $CD34^+CD38^-$ cells, $CD34^+$Thy-I cells, $CD34^+$Thy-I$CD38^-$ cells, $CD33^+$ cells, stem cells, progenitor cells, long-term culture initiating cells (LTC-IC), cells that fulfill self renewal potential, cells that fulfill pluripotential characteristics, cells that initiate long term bone marrow culture.

Determination of amounts of growth hormone, of one of its derivatives or of any factor inducing growth hormone release to be administered in a method or use of the invention described above is within the skill of the art.

Typical dosage of growth hormone, of one of its derivatives or of any factor inducing growth hormone release will start at about 1 microgram per kilogram of the patient weight per day and dose will be escalated until the desired effect (hematopoietic recovery or engraftment) is reached.

The dosage of growth hormone, of one of its derivatives or of any factor inducing growth hormone release administered depends upon the age, sex, health

and weight of the donor, type of previous or concurrent treatment, if any, frequency of the treatment and the nature of the effect desired.

Growth hormone or its derivatives or any factor inducing growth hormone release may be administered alone or in conjunction or association with other factors.

Growth hormone or its derivatives or any factor inducing growth hormone release may advantageously be present in a composition or a medicament which comprises further one or several compound(s) chosen among the compounds belonging to the following groups : hematopoietic growth factors, cytokines, chemokines, monoclonal antibodies.

Growth hormone or its derivatives or any factor inducing growth hormone release and one or several compound(s) chosen among the compounds belonging to the following groups : hematopoietic growth factors, cytokines, chemokines, monoclonal antibodies can be administered simultaneously or at different times and/or at the same site or at different site(s) and/or in the same or in a different composition or medicament.

The cytokine group can comprise IL-1,IL-3, G-CSF, GM-CSF or SCF. The chemokine group can comprise MIP-1α or thrombopoietin (TPO). The monoclonal antibody group can comprise anti-VLA-4 antibodies.

Preferably, Growth hormone or its derivatives or any factor inducing growth hormone is present in a composition or a medicament which comprises granulocyte colony stimulating factor (G-CSF).

Preferably, Growth hormone or its derivatives or any factor inducing growth hormone is associated with G-CSF.

Growth hormone or its derivatives or any factor inducing growth hormone and G-CSF can be administered simultaneously or at different times and/or at the same

site or at different site(s) and/or in the same or in a different composition or medicament.

Growth hormone or its derivatives or any factor inducing growth hormone release and G-CSF may advantageously be administered separately.

Growth hormone or its derivatives or any factor inducing growth hormone release and/or G-CSF are administered in an amount sufficient to enhance hematopoietic reconstitution or engraftment.

Administration in an amount sufficient to enhance hematopoietic reconstitution or engraftment can mean one or several administration(s), one or several times a day and during one or several days for a cumulated amount sufficient to enhance hematopoietic reconstitution or engraftment.

The pharmaceutical compositions or medicaments or compositions which are used in the methods and uses of the invention are in a pharmaceutical acceptable form optionally combined with an acceptable carrier.

These compositions or medicaments can be administered by any means that achieve their intended purposes.

The compositions or medicaments used in the methods or uses of the invention may be administered alone or in conjunction with other therapeutics directed to a disease or directed to other symptoms thereof.

The compositions or medicaments used in the methods or uses of the invention may be administered by the intravenous or the subcutaneous route.

The compositions or medicaments used in the methods or uses of the invention may be administered by parenteral routes such as subcutaneous, intravenous, intramuscular, intraperitoneal, or transdermal route or by mucosal routes such as buccal or oral route.

The composition or medicament comprising growth hormone or one of its derivatives or any factor inducing growth hormone release may be administered by parenteral routes such as subcutaneous, intravenous, intramuscular, intraperitoneal, or transdermal route or by mucosal routes such as buccal or oral route.

Preferably, growth hormone or one of its derivatives or any factor inducing growth hormone release is administered subcutaneously.

The total dose or amount required for each treatment, method or use of the invention may be administered in multiple or single dose.

The composition or medicament comprising growth hormone or one of its derivatives or any factor inducing growth hormone release may be administered daily or three times a day.

Preferably, the composition or medicament comprising growth hormone or one of its derivatives or any factor inducing growth hormone release is administered three times a day.

Preferably, growth hormone or one of its derivatives or any factor inducing growth hormone release is administered daily or three times a day.

In a preferred embodiment, growth hormone or one of its derivatives or any factor inducing growth hormone release is administered three times a day.

If a method or use of the invention comprises the administration of growth hormone or one of its derivatives or any factor inducing growth hormone release and G-CSF, G-CSF is preferably administered once a day and/or subcutaneously.

If a method or use of the invention comprises the administration of growth hormone or one of its derivatives or any factor inducing growth hormone release and G-CSF, growth hormone or one of its derivatives or

any factor inducing growth hormone release is preferably administered three times a day and G-CSF is preferably administered daily.

The administration of the medicament may be made over a period of 3 days along, until leukapheresis or until full recovery.

The administration of the medicament may be made from day 1 to day 3 after transplantation.

The term « transplantation » encompasses bone marrow transplantation or hematopoietic stem cells transplantation.

The composition or medicament comprising growth hormone or one of its derivatives or any factor inducing growth hormone release may be a daily administration that can start up to 20 days pre-leukapheresis.

The composition comprising growth hormone or one of its derivatives or any factor inducing growth hormone release may be administered over a period of 5 days or over a period of 10 days until the desired effect (hematopoietic recovery or engraftment) is reached.

Preferably, the composition comprising growth hormone or one of its derivatives or any factor inducing growth hormone release is administered until the desired effect (hematopoietic recovery or engraftment) is reached.

Methods and uses of the invention are advantageously carried out after chemotherapy, radiotherapy, myelosuppressive therapy, transplantation or engraftment of cells capable of regenerating hematopoiesis in vivo or transplantation of bone-marrow.

Methods and uses of the invention are advantageously carried out around 7 days after the beginning of a chemotherapeutic treatment or around 2 days after the end of a chemotherapeutic treatment.

In a preferred embodiment, growth hormone or one of its derivatives or any factor inducing growth hormone release and G-CSF are administered until hematopoietic reconstitution or engraftment. In this preferred embodiment, growth hormone or one of its derivatives or any factor inducing growth hormone release is preferably administered three times a day and G-CSF is preferably administered once a day.

Growth hormone used in the medicament may advantageously be recombinant growth hormone.

Growth hormone used in the medicament may advantageously be human growth hormone.

In a third part, the invention provides a combination of the methods and uses of the first part of the invention (mobilisation) with the methods and uses of the second part of the invention (recovery).

Said combination methods and uses are mobilisation and recovery methods which may be applied in cases autologous transplantation of hematopoietic stem cells wherein the donor and the recipient is the same person or individual. Thus, growth hormone or one of its derivatives or any factor inducing growth hormone release can be used as a mobilising agent in a first mobilisation step which is a pre-treatment in view of blood cells extraction and as an hematopoietic recovery agent in a second step following transplantation.

Said combination methods and uses are very useful. In fact, transplantation of cells mobilized by growth hormone or one of its derivatives or any factor inducing growth hormone release to a patient results in faster haematological recovery than transplantation without a prior mobilization treatment of said patient.

Methods and uses of the invention can be applied in many clinically important fields, namely autologous bone marrow transplantation, allogeneic bone marrow

transplantation, gene therapy, hematopoietic stem cells transplantation, transplantation of cells capable of regenerating hematopoiesis in vivo, radiotherapy, chemotherapy, myelosuppressive or myelotoxic therapy.

Methods and uses of the invention can be applied to treat a patient who has received radiotherapy or chemotherapy, who has been transplanted with bone-marrow or cells capable of regenerating hematopoiesis in vivo, or who has received myelotoxic or myeloablative therapy.

LEGEND OF THE FIGURES :

Figure 1 :

Abbreviations :

- **GH** : Growth Hormone
- **G-CSF** : Granulocyte-Cell Stimulating Factor
- **ND** : Not Detectable

Figure 2 :

This graph depicts the number of $CD34^+$ cells/μl of blood obtained in a patient during 3 cycles of chemotherapy after a mobilization treatment with G-CSF alone (cycle 1), GH + G-CSF (cycle 2) and G-CSF alone (cycle 3).

Examples

Abbreviations and notes :

- **BFU-E** : burst forming unit, erythroid
- **CFU-C** : colony forming unit, culture
- **CFU-GM** : colony forming unit, granulocyte and macrophage
- **CFU-Meg** : colony forming unit, megakaryocyte
- **G-CSF** : granulocyte colony stimulating factor
- **IGF-I** : insulin growth factor I
- **LTC-IC** : long term culture initiating cell
- **HGH** : human growth hormone

- **RhG-CSF** : recombinant human granulocyte colony stimulating factor
- **RhGH** : recombinant human growth hormone

Example 1 : Mobilization activity of hGH studied in a murine preclinical model

BALB/c mice are given 10µL intraperitoneal injections of rhGH every day for total of 10 days. The total CFU-C or BFU-E activity circulating in the peripheral blood on day 5 and day 10, respectively, is determined according to standard in vitro culture techniques, and compared with :

(i) steady-state pretreatment levels,

(ii) absolute CFU-C and BFU-E counts on day 3 and day 5, respectively following treatment with rhG-CSF given intraperitoneally at 10µL every day for 5 consecutive days.

Example 2 : Selection criteria for the mobilization and recovery clinical studies

A) Inclusion criteria :

- Written informed consent
- Age 18 years and 60 years
- Histologically confirmed high-risk cancer (lymphome cancer) undergoing high-dose chemotherapy according to current INT guidelines.

B) Exclusion criteria :

- Patients heavily pretreated with chemotherapy courses of chemotherapy) and/or radiotherapy.

- Renal (creatinine > 1,5 N), or hepatic insufficience and/or SGPT > 2,5 N ; bilirubin > 1,5 N), or severe CNS or psychiatric disease.
- Clinically significant cardiac disease or myocardiac. Left ventricular ejection fraction < 50% at rest by echocardiography assessment or < 55% by isotopic measurement.
- Hepatitis B or C, or HIV test positive.

Example 3 : Baseline study procedure for the mobilization and recovery clinical studies

Several parameters are studied during the mobilisation and recovery clinical studies :

- Complete medical history, physical examination, cardiac examination, left ventricular ejection fraction (LVEF) by multigated scintigraphic scan or echography, chest X-ray
- Pregnancy test (if applicable)
- HBV, HCV and HIV test
- Complete blood count with differential
- Absolute counts of circulating $CD34^+$ cells and CFU
- Blood chemistry (transaminases, serum phosphatase, gammaGT, LDH, total bilirubin, BUN, creatinine, glycemia, Na, K, Ca, P, uric acid, total protein, albumin, cholesterol, triglycerides
- Bilateral bone marrow biopsy
- Informed consent

Example 4 : Main parameters of toxicity for the mobilization and recovery clinical studies

- Tumor growth (mobilization study only)
- Clinical and instrumental symptoms
- Laboratory tests for cardiac, liver and renal function

Example 5 : Mobilization clinical study

A) Objectives of the mobilization clinical study

- To assess the activity of rhGH in :
  (i) increasing circulating $CD34^+$ cells, and
  (ii) expanding the bone marrow hematopoietic compartment, so to allow an enhanced mobilization by subsequent rhG-CSF administration
- To assess the safety and tolerability of rhGH, given with rhG-CSF to cancer patients following chemotherapy (hematologic recovery study).

B) Treatment plan

Mobilization study with rhGH :

- rhGH is administered from day 1 to 10 by the intravenous route. Dosage of rhGH is started at about 1 micrograms per kilogram of the patient weight per day and dose will be escalated until the desired effect (mobilization or peripheralisation of circulating cells capable of regenerating hematopoiesis in vivo, increase in said donor of the number of circulating cells capable of regenerating hematopoiesis in vivo, reduction of the number of leukapheresis required to collect sufficient amount of circulating cells for transplantation, reduction of the volume of blood required to be processed in order to obtain the specified target number of circulating

cells capable of regenerating hematopoiesis in vivo) is reached.xx μg/kg QD, iv)

Mobilization study with rhGH and rhG-CSF :

- rhGH administration : rhGH is administered from day 1 to 5 by the intravenous route. Dosage of rhGH is started at about 1 micrograms per kilogram of the patient weight per day and dose will be escalated until the desired effect (mobilization or peripheralisation of circulating cells capable of regenerating hematopoiesis in vivo, increase in said donor of the number of circulating cells capable of regenerating hematopoiesis in vivo, reduction of the number of leukapheresis required to collect sufficient amount of circulating cells for transplantation, reduction of the volume of blood required to be processed in order to obtain the specified target number of circulating cells capable of regenerating hematopoiesis in vivo) is reached.
- rhG-CSF administration (10 μg/kg QD, iv) from completion of $CD34^+$ cell harvest (target cell dose is 8 X $10e^6$ $CD34^+$ cells/kg body weight).

C) Main parameters of activity

Starting from day +6, the following parameters are assessed :

- Absolute $CD34^+$ cell counts/μL (daily in the periphery and once in the leukapheresed cells)
- Absolute CFU-GM counts/μL (daily in the periphery and once in the leukapheresed cells)

D) Study procedure

- Daily assessment of $CD34^+$ cells/μL and CFU-GM peripheral blood, from day +5 until leukapheresis.
- Total yield of $CD34^+$ cells, CFU-GM, BFU-E, CFU IC in leukapheresed cells.

- Toxicity assess through clinical and in examinations (EKG, chest X-ray, and other examinations as required).
- Measurement and evaluation of all tumor parameters ending the mobilization study

Example 6 : Recovery clinical study

A) Objectives of the recovery clinical study

- To assess the ability of rhGH, given alone or in combination to hasten the recovery of WBC, RBC and platelets in the peripheral blood of cancer patients treated with high-dose chemotherapy and peripheral blood stem-cell autografting.
- To assess the safety and tolerability of rhGH, given with rhG-CSF to cancer patients following chemotherapy.

B) Treatment plan

- Administration of high-dose chemotherapy, followed by infusion on day 0 of an optimal amount (i.e. $8X10e^6$ $CD34^+$ cells/kg) of crypreserved cells harvested in the mobilization study.
- Co-administration (iv) of rhGH and rhG-CSF μg/kg QD, iv) from day 1 until stable (i.e. for three consecutive days) recovery of granulocytes above 1000μL, and platelet counts above 50,000/μL.

C) Main parameters of activity

Starting from day +0, and until full and stable recovery, the following parameters will be assessed :

- Absolute granulocyte counts/μL (daily)
- Absolute platelet counts/μL (daily)
- Absolute erythrocyte counts/μL (daily)
- Granulocyte nadir

- Platelet nadir
- Extent and duration of neutropenia
- Extent and duration of thrombocytopenia
- Extent and duration of hematopoietic support tranfusions, RBC transfusions)
- Duration of infectious prophylaxis, and infections
- Hemorrhages

D) Study procedure

- Daily assessment of WBC, RBC and platelet count
- Number of platelet transfusions
- Number of RBC transfusions
- Type and severity of fever and documented infection
- Clinical and instrumental assessment of toxicities hematological

Example 7 : Results of the mobilization clinical studies

I - Studies of mobilization within 2 cycles of chemotherapy

A) Treatment plan

Three patients with relapsed Hodgkin's disease have received the following two cycles of treatment :

- cycle 1(control cycle) :
- Ifosfamide (agent for chemotherapy) : 3 g/m^2 iv (intravenous) (once a day), day 1-4 ;
- Vinorelbine (agent for chemotherapy) : 25 mg/m^2 iv (once a day), day 1 and 5 ;
- G-CSF : 5 $\mu g/kg$ sc (subcutaneous) (once a day), from day 7 until leukapheresis or until recovery of a sufficient number of CD34+ cells(around 3 to 8-10^6 cells/kg).

- cycle 2 :
- Ifosfamide : 3 g/m2 iv (once a day), day 1-4 ;
- Vinorelbine : 25 mg/m^2 iv (once a day), day 1 and 5 ;
- G-CSF : 5 µg/kg sc (once a day) from day 7 until recovery of a sufficient number of CD34+ cells (around 3 to 8-10^6 cells/kg) or until leukapheresis ;
- rhGH(recombinant human Growth Hormone) : 33 µg/kg sc (three times a day), from day 7 until recovery of a sufficient number of CD34+ cells (around 3 to 8-10^6 cells/kg)or until leukapheresis.

B)Results

The results are depicted in the table of figure 1.

No toxicity was observed except hyperglycemia requiring insulin administration.

As compared with the control (cycle 1), the addition of rhGH in cycle 2 resulted in :

1) Doubling or tripling the mobilization of CD34$^+$ cells in the blood stream
2) Recuperation of leukapheresed CD34$^+$ cells or increasing of the number of CD34$^+$ leukapheresed cells.

The increase of the number of CD34$^+$ leukapheresed cells induced by GH allows the harvest from all three patients of an amount of CD34+ cells adequate for autologous transplantation (around 3 to 8-10^6 cells/kg).

II - Studies of mobilization within 3 cycles of chemotherapy

A) Treatment plan

One patient with relapsed Hodgkin's disease has received the following 3 cycles of treatment :

- cycle 1 :
- Ifosfamide (agent for chemotherapy) : 3 g/m² iv (intravenous) (once a day), day 1-4 ;
- Vinorelbine (agent for chemotherapy) : 25 mg/m² iv (once a day), day 1 and 5 ;
- G-CSF : 5 µg/kg sc (subcutaneous) (once a day), from day 7 until leukapheresis or recovery of a sufficient amount of CD34+ cells (around 3 to 8-10^6 cells/kg).

- cycle 2 :
- Ifosfamide : 3 g/m2 iv (once a day), day 1-4 ;
- Vinorelbine : 25 mg/m² iv (once a day), day 1 and 5 ;
- G-CSF : 5 µg/kg sc (once a day) from day 7 until recovery of a sufficient amount of CD34+ cells (around 3 to 8-10^6 cells/kg) or until leukapheresis ;
- rhGH (recombinant human Growth Hormone) : 33 µg/kg sc (three times a day), from day 7 until recovery of a sufficient amount of CD34+ cells (around 3 to 8-10^6 cells/kg) or until leukapheresis.

- cycle 3 :
- Ifosfamide (agent for chemotherapy) : 3 g/m² iv (intravenous) (once a day), day 1-4 ;
- Vinorelbine (agent for chemotherapy) : 25 mg/m² iv (once a day), day 1 and 5 ;
- G-CSF : 5 µg/kg sc (subcutaneous) (once a day), from day 7 until leukapheresis or until recovery of a

sufficient amount of CD34+ cells(around 3 to 8-10^6 cells/kg).

B) Results

The results of the clinical treatment recited in section A above are depicted in the graph of Figure 2.

Graph of figure 2 shows the time course of $CD34^+$ cell mobilization, following three consecutive chemotherapy cycles beginning every 21 days.

Each point depicted in the graph of figure 2 corresponds to the measure of the number of $CD34^+$ cells/μl of blood found in a sample of blood of 1 milliliter.

The results show that cycle 2 (addition of rhGH) is clearly superior to cycles 1 and 3. Thus, the mobilization or $CD34^+$ in the blood is enhanced by the addition of rhGH.

Enhancement of mobilization of $CD34^+$ cells in the blood by GH is high, especially since the patient studied receives several courses of myelotoxic chemotherapy, and since each susbsequent course hampers the extent of mobilization. The declining numbers of circulating $CD34^+$ cells after consecutive myelotoxic chemotherapy and mobilization cycles can be observed by comparing cycle 1 and cycle 3.

The blood of the patient is leukapheresed when the number of $CD34^+$ cells/μl of blood measured is maximal (day 13 of cycle 1 ; day 20 of cycle 2).

The leukapheresed cells are cryopreserved and will be reinfused in the patient after a myeloablative therapy.

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CLAIMS

1. Method of preparation of a population of circulating cells capable of regenerating hematopoiesis in vivo comprising :

a) administering to a donor a composition comprising growth hormone or one of its derivatives or any factor inducing growth hormone release in an amount sufficient to increase in said donor the number of circulating cells capable of regenerating hematopoiesis in vivo.

b) isolating a population of circulating cells capable of regenerating hematopoiesis in vivo from the peripheral blood of said donor.

2. Method of preparation of a donor of circulating cells, which cells are capable of regenerating hematopoiesis in vivo comprising the administration to said donor of a composition comprising growth hormone or one of its derivatives or any factor inducing the growth hormone release in an amount sufficient to increase the number of circulating cells capable of regenerating hematopoiesis in vivo in said donor.

3. Method for increasing the number of circulating cells capable of regenerating hematopoiesis in vivo in a donor by administration of a composition comprising the growth hormone or one of its derivatives or any factor inducing the growth hormone release to said donor.

4. Method according to any one of the preceding claims wherein the circulating cells capable of regenerating hematopoiesis in vivo are $CD34^+$ cells.

5. Method according to claim 4 wherein the increased number of $CD34^+$ cells is more than 10, 25, 34 or 80 $CD34^+$ cells per microliter of peripheral blood.

6. Method according to any one of the preceding claims wherein the increased number of $CD34^+$ cells is at least $2x10^6$ $CD34^+$ cells per kilogram of recipient body weight, or at least $4x10^6$ $CD34^+$ cells per kilogram of recipient body weight or at least $8x10^6$ $CD34^+$ cells per kilogram of recipient body weight.

7. Method according to any one of the preceding claims wherein the increased number of $CD34^+$ cells is at least $2x10^6$, $4x10^6$, $5x10^6$, $6x10^6$, $8x10^6$, $15x10^6$ $CD34^+$ cells per kilogram of donor body weight.

8. Method according to any one of the preceding claims wherein the increased number of circulating cells capable of regenerating hematopoiesis in vivo corresponds to around or more than $1x10^5$ GM-CFC per kilogram of donor or recipient body weight.

9. Method according to any one of the preceding claims wherein the increased number of circulating cells capable of regenerating hematopoiesis in vivo corresponds to around or more than 500 CFU-GM per milliliter of peripheral blood.

10. Method according to to any one of the preceding claims wherein the increased number of circulating cells capable of regenerating hematopoiesis in vivo corresponds to an increased level of CFU-C, CFU-Meg or BFU-E.

11. Method according to any one of the preceding claims wherein the increased number of circulating cells capable of regenerating hematopoiesis in vivo substantially corresponds to a white blood cell count of around or more than 1000 cells per microliter of peripheral blood.

12. Method according to any one of the preceding claims wherein the circulating cells capable of regenerating hematopoiesis in vivo are $CD34^+/CD33^+$ cells and/or $CD34^+/CD38^-$ cells and/or $CD34^+$/Thy-I cells and/or $CD34^+$/Thy-I/$CD38^-$ cells and/or $CD33^+$ cells and/or bone-

marrow stem cells and/or progenitor cells and/or long-term culture initiating cells (LTC-IC) and/or cells that fulfill self renewal potential and/or cells that fulfill pluripotential characteristics and/or cells that initiate long term bone marrow culture and/or cells that can generate multiple cell lineages.

13. Method of preparation of a population of circulating cells capable of regenerating hematopoiesis in vivo comprising :

a) administering to a donor a composition comprising growth hormone or one of its derivatives or any factor inducing growth hormone release in an amount sufficient to reduce the volume of blood required to be processed in order to obtain the specified target number of circulating cells capable of regenerating hematopoiesis in vivo.
b) isolating said reduced volume of blood
c) isolating a population of circulating cells capable of regenerating hematopoiesis in vivo from said isolated volume.

14. Method of preparation of a donor of circulating cells, which cells are capable of regenerating hematopoiesis in vivo comprising the administration to said donor of a composition comprising growth hormone or one of its derivatives or any factor inducing growth hormone release in an amount sufficient to reduce the volume of blood required to be processed in order to obtain the specified target number of circulating cells capable of regenerating hematopoiesis in vivo and/or to reduce the number of leukapheresis required to collect sufficient amount of circulating cells capable of regenerating hematopoiesis in vivo to be tranplanted.

15. Method for reducing the volume of blood required to be processed in order to obtain the specified target number of circulating cells capable of

regenerating hematopoiesis in vivo in a donor and/or for reducing the number of leukapheresis required to collect sufficient amount of circulating cells capable of regenerating hematopoiesis in vivo to be tranplanted by administration of a composition comprising the growth hormone or one of its derivatives or any factor inducing the growth hormone release to said donor .

16. Method according to any one of the preceding claims wherein the specified target number of circulating cells capable of regenerating hematopoiesis in vivo is at least $2x10^4$ LTC-IC per kg of donor or recipient body, around or more than $2x10^6$ $CD34^+$ cells per kilogram of donor or recipient body weight, around or more than $4x10^6$ $CD34^+$ cells per kilogram of donor or recipient body weight or around or more than $8x10^6$ $CD34^+$ cells per kilogram of donor or recipient body weight.

17. Method according to any one of the preceding claims wherein the required volume of blood is comprised in a range of about 30 to about 900 milliliters.

18. Method according to any one of the preceding claims wherein the composition comprises further one or several compound chosen among the following groups of compounds : hematopoietic growth factors, cytokines, chemokines, monoclonal antibodies.

19. Method according to any one of the preceding claims wherein the cytokine group comprises IL-1,IL-3, G-CSF, GM-CSF or SCF ; the chemokine group comprises MIP-1α or thrombopoietin (TPO) ; the monoclonal antibody group comprises anti-VLA-4 antibodies.

20. Method according to any one of the preceding claims wherein the composition comprises Growth Hormone and G-CSF.

21. Method according to any one of the preceding claims wherein growth-hormone is administered in an amount comprised between 20 to 50 µg/kg of body weight,

in an amount comprised between 30 to 40 µg/kg of body weight or in an amount of around 33 µg per kilogram of body weight.

22. Method according to claims 19 to 21 wherein the G-CSF is administered in an amount comprised between 3 to 15 µg/kg of body weight, in an amount comprised between 4 to 12 µg/kg of body weight or in an amount of around 5 or 10 µg per kilogram of body weight.

23. Method according to any one of the preceding claims wherein the administration of Growth Hormone is made three times a day and the administration of G-CSF is made daily.

24. Method according to any one of the preceding claims wherein the administration is made by parenteral, subcutaneous, intravenous, intramuscular, intraperitoneal, transdermal or buccal routes.

25. Method according to any one of the preceding previous claims wherein the administration is daily or three times a day.

26. Method according to any one of the preceding claims wherein the administration is made over a period of 5 days, over a period of 10 days, until leukapheresis, until mobilization or peripheralisation of circulating cells capable of regenerating hematopoiesis in vivo, until increase of the number of circulating cells capable of regenerating hematopoiesis in vivo, until reduction of the number of leukapheresis required to collect sufficient amount of circulating cells for transplantation, and/or until reduction of the volume of blood required to be processed in order to obtain the specified target number of circulating cells capable of regenerating hematopoiesis in vivo.

27. Method according to any one of the preceding claims wherein the administration(s) is/are made after

chemotherapy, radiotherapy, myelotoxic or myelosuppressive therapy, transplantation of cells capable of regenerating hematopoiesis in vivo or bone-marrow transplantation.

28. Method according to claim 27 wherein the administration(s) begin(s) around 7 days after the beginning of a chemotherapeutic treatment or around 2 days after the end of a chemotherapeutic treatment.

29. Method according to any one of the preceding claims wherein the growth hormone is recombinant growth hormone.

30. Method according to any one of the preceding claims wherein the growth hormone is human growth hormone.

31. Use of human growth hormone or one of its derivatives or any factor inducing human growth hormone release to prepare a medicament to increase the peripheral White Blood Cell count and/or granulocytes count and/or lymphocytes count and/or erythrocyte count in a human being.

32. Use of human growth hormone or one of its derivatives or any factor inducing human growth hormone release to prepare a medicament to increase the peripheral White Blood Cell count and/or granulocytes count and/or lymphocytes count and/or erythrocyte count following bone marrow transplantation, transplantation of cells capable of regenerating hematopoiesis in vivo, radiotherapy, chemotherapy, myelotoxic therapy or myelosuppressive therapy in a human being.

33. Use of human growth hormone or one of its derivatives or any factor inducing human growth hormone release to prepare a medicament to increase the peripheral White Blood Cell (WBC) count and/or granulocytes count and/or lymphocytes count and/or erythrocyte count in a human being after transplantation,

radiotherapy, chemotherapy, myelotoxic therapy or myelosuppressive therapy.

34. Use according to any one of claims 31 to 33 wherein growth hormone is human growth hormone.

35. Use of growth hormone or one of its derivatives or any factor inducing growth hormone release to prepare a medicament for reducing the period of time necessary to recover a normal peripheral White Blood Cell (WBC) count and/or granulocytes count and/or neutrophil count and/or lymphocytes count and/or platelet count and/or erythrocyte count.

36. Use according to claim 35 wherein the normal neutrophil count is at least $0.5x10^9$ per liter of peripheral blood.

37. Use according to claim 35 wherein the normal platelet counts is at least $20x10^9$ per liter of peripheral blood.

38. Use of growth hormone or one of its derivatives or any factor inducing growth hormone release to prepare a medicament for reducing the duration of neutropenia and/or thrombocytopenia and/or anemia and/or hemorrhages and/or duration of prophylaxis.

39. Use of growth hormone or one of its derivatives or any factor inducing growth hormone release to prepare a medicament for reducing the duration and/or severity of fever and infections after transplantation, radiotherapy, chemotherapy or myelosuppresive therapy.

40. Use of growth hormone or one of its derivatives or any factor inducing growth hormone release to prepare a medicament for recovering a granulocytes count which is at least 1000 per microliter of peripheral blood after transplantation, radiotherapy, chemotherapy or myelosuppresive therapy.

41. Use of growth hormone or one of its derivatives or any factor inducing growth hormone release to prepare

a medicament for recovering a platelet count which is at least 50,000 per microliter of peripheral blood after transplantation, radiotherapy, chemotherapy or myelosuppresive therapy.

42. Use of growth hormone or one of its derivatives or any factor inducing growth hormone release to prepare a medicament for treating a neoplastic disease, an hematological disorder, malignancies, congenitally or genetically determined hematopoietic abnormalities, aplastic anemia, leukemia and/or osteopetrosis.

43. Use of growth hormone or one of its derivatives or any factor inducing growth hormone release to prepare a medicament for preventing and/or treating opportunistic infections after transplantation or for limiting the risk of tumor recurrence after transplantation.

44. Use of growth hormone or one of its derivatives or any factor inducing growth hormone release to prepare a medicament for preventing or treating anemia and/or neutropenia following radiotherapy and/or chemotherapy and /or hematopoietic stem cells transplantation and/or transplantation of cells capable of regenerating hematopoiesis and/or Bone marrow transplantation.

45. Use according to any one of claims 31-44 wherein the medicament comprises further one or several compound(s) chosen among the following groups of compounds : hematopoietic growth factors, cytokines, chemokines, monoclonal antibodies.

46. Use according to claim 45 wherein the cytokines group comprises IL-1,IL-3, IL-6, IL-11, Insulin-like growth factor 1 (IGF-1), G-CSF, GM-CSF or SCF ; the chemokines group comprises MIP-1α or thrombopoietin (TPO) ; the monoclonal antibodies group comprises anti-VLA-4 antibodies.

47. Use according to any one of claims 31-46 wherein the medicament comprises Growth Hormone and G-CSF.

48. Use according to any one of claims 31-47 wherein the administration is made by parenteral, subcutaneous, intravenous, intramuscular, intraperitoneal, transdermal or buccal routes.

49. Use according to any one of claims 31-48 wherein the administration is daily or three times a day.

50. Use according to any one of claims 45-49, wherein the administration of growth hormone is made three times a day and the administration of G-CSF is daily.

51. Use according to any one of claims 31-50 wherein the administration is made over a period of 3 days, until leukapheresis, until hematopoietic reconstitution or until engraftment.

52. Use according to any one of the preceding claims wherein the administration(s) is/are made after chemotherapy, radiotherapy, myelotoxic or myelosuppressive therapy, transplantation of cells capable of regenerating hematopoiesis in vivo or bone-marrow transplantation.

53. Use according to claim 52 wherein the administration(s) begin(s) around 7 days after the beginning of a chemotherapeutic treatment or around 2 days after the end of a chemotherapeutic treatment.

54. Use according to any one of claims 31-53 wherein the administration is made from day 1 to day 3 after transplantation.

55. Use according to any one of claims 39 to 41, 43 or 54 wherein the transplantation is bone marrow transplantation or hematopoietic stem cells transplantation.

56. Use according to any one of claims 31-55 wherein growth hormone is recombinant growth hormone.

57. Use according to any one of claims 31-56 wherein growth hormone is human growth hormone.

58. Use according to any one of claims 31 to 58 which is preceded in time by a method according to any one of claims 1 to 30.

59. Method for the enhancement of hematopoiesis reconstitution in a human being comprising the steps of :

a) administering to a donor a composition comprising growth hormone or one of its derivatives or any factor inducing growth hormone release in an amount sufficient to increase in said donor the number of circulating cells capable of regenerating hematopoiesis in vivo.

b) isolating a population of circulating cells capable of regenerating hematopoiesis in vivo from the peripheral blood of said donor.

c) transplantation of the cells recovered in step (b) to an individual, and

d) administration of growth hormone or one of its derivatives or any factor inducing growth hormone release in an amount sufficient to accelerate hematopoietic recovery.

60. Method for the enhancement of hematopoiesis reconstitution in a human being comprising the steps of :

a) administering to a donor a composition comprising growth hormone or one of its derivatives or any factor inducing growth hormone release, and G-CSF in an amount sufficient to increase in said donor the number of circulating cells capable of regenerating hematopoiesis in vivo.

b) isolating a population of circulating cells capable of regenerating hematopoiesis in vivo from the peripheral blood of said donor.

c) transplantation of the cells recovered in step (b) to an individual, and

d) administration of growth hormone or one of its derivatives or any factor inducing growth hormone release and G-CSF in an amount sufficient to accelerate hematopoietic recovery.

Agents administered after chemotherapy		Absolute peak number of circulating $CD34^+/\mu L$			Total amount of leukapheresed $CD34^+$ cells/kg of body weight		
		Patient 1	Patient 2	Patient 3	Patient 1	Patient 2	Patient 3
G-CSF	Cycle 1	10	12	50	ND	ND	5.10^6
GH + G-CSF	Cycle 2	25	34	80	5.10^6	$14.9\ 10^6$	$6.1\ 10^6$

Figure 1

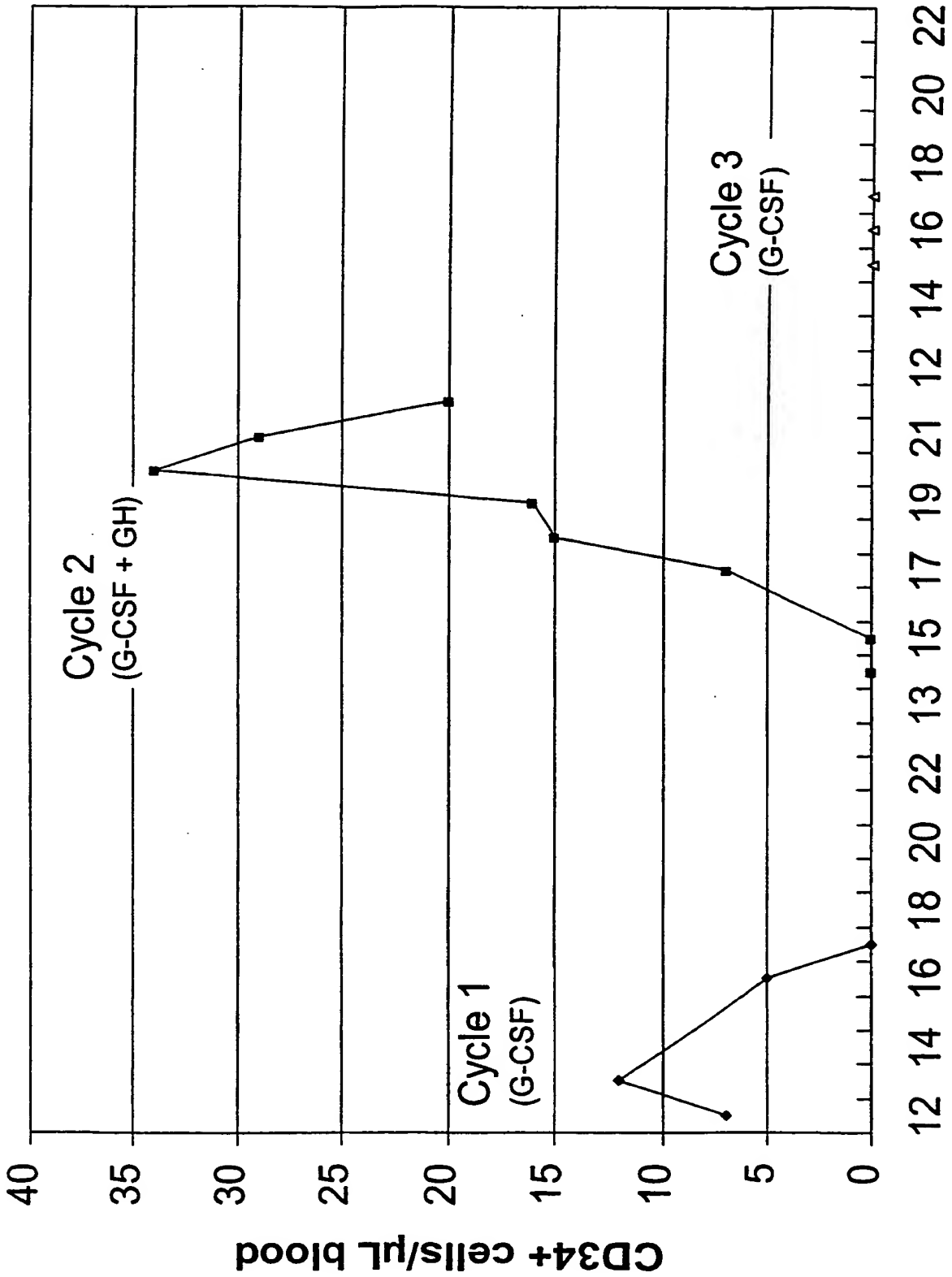
CD34+ cells/µL blood
0
5
10
15
20
25
30
35
40
12 14 16 18 20 22 13 15 17 19 21 12 14 16 18 20 22
Days post-therapy
Cycle 1
(G-CSF)
Cycle 2
(G-CSF + GH)
Cycle 3
(G-CSF)

Figure 2

INTERNATIONAL SEARCH REPORT

Int. Application No
PCT/EP 99/10470

A. CLASSIFICATION OF SUBJECT MATTER
IPC 7 A61K38/27 C12N5/06 C12N5/08 A61K35/14 A61K35/28
//(A61K38/27,38:19)

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
IPC 7 A61K C12N C07K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	PATENT ABSTRACTS OF JAPAN vol. 095, no. 007, 31 August 1995 (1995-08-31) & JP 07 101877 A (MITSUI TOATSU CHEM INC), 18 April 1995 (1995-04-18) abstract	35, 37-39, 41-43, 48,49, 52-55, 57,58
X	TIAN Z G ET AL: "Recombinant human growth hormone promotes hematopoietic reconstitution after syngeneic bone marrow transplantation in mice." STEM CELLS, (1998) 16 (3) 193-9., XP002114334	1-21, 23-60
Y	the whole document	22

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[X] Further documents are listed in the continuation of box C.

[X] Patent family members are listed in annex.

* Special categories of cited documents :

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier document but published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

"&" document member of the same patent family

Date of the actual completion of the international search	Date of mailing of the international search report
29 February 2000	15/03/2000

Name and mailing address of the ISA	Authorized officer
European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Tx. 31 651 epo nl, Fax: (+31-70) 340-3016	Stein, A

Form PCT/ISA/210 (second sheet) (July 1992)

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category °	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	BREGNI M ET AL: "Comparative effects of granulocyte-macrophage colony-stimulating factor and granulocyte colony - stimulating factor after high-dose cyclophosphamide cancer therapy." JOURNAL OF CLINICAL ONCOLOGY, (1996 FEB) 14 (2) 628-35. , XP000877223 the whole document	22
X	MURPHY W J ET AL: "GROWTH HORMONE EXERTS HEMATOPOIETIC GROWTH-PROMOTING EFFECTS IN VIVO AND PARTIALLY COUNTERACTS THE MYELOSUPPRESSIVE EFFECTS OF AZIDOTHYMIDINE" BLOOD, vol. 80, no. 6, 15 September 1992 (1992-09-15), pages 1443-1447, XP002065327 the whole document	1-60
X	MIYASHITA Y: "The effect of growth hormone on leukopoiesis: in vivo and in vitro studies." NIPPON NAIBUNPI GAKKAI ZASSHI. FOLIA ENDOCRINOLOGICA JAPONICA, (1991 JUL 20) 67 (7) 785-95., XP002114569 abstract	1-60
X	MURPHY W J ET AL: "Human growth hormone promotes engraftment of murine or human T cells in severe combined immunodeficient mice." PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES OF THE UNITED STATES OF AMERICA, (1992 MAY 15) 89 (10) 4481-5., XP002114570 the whole document	2-12, 18-20, 24-27, 29, 31-33, 35-39, 42-44, 48,49, 51,52, 54-56,58
A	US 5 199 942 A (GILLIS STEVEN) 6 April 1993 (1993-04-06) column 2, line 6 - line 36 column 3, line 56 -column 4, line 66 column 5, line 16 - line 23 column 5, line 53 - line 62 column 5, line 53 - line 62; examples 2-8	1-60
A	KÖRBLING M ET AL: "Allogeneic peripheral blood stem cell transplantation using normal patient-related pediatric donors" BONE MARROW TRANSPLANTATION, vol. 18, no. 5, November 1996 (1996-11), pages 885-890, XP002114335 the whole document	19,20, 22-26, 46-51,60

-/—

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category °	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	OHMIZONO Y ET AL: "THROMBOPOIETIN AUGMENTS EX VIVO EXPANSION OF HUMAN CORD BLOOD -DERIVED HEMATOPOIETIC PROGENITORS IN COMBINATION WITH STEM CELL FACTOR AND FLT3 LIGAND" LEUKEMIA, vol. 11, no. 4, 1 April 1997 (1997-04-01), pages 524-530, XP002053902 the whole document	1-26, 45-48, 55,57,58
A	WO 94 28916 A (BRITISH BIOTECH PHARM ;COMER MICHAEL BERISFORD (GB); MCCOURT MATTH) 22 December 1994 (1994-12-22) page 5, line 25 -page 6, line 21; examples 2-8 page 11, line 22 -page 12, line 4; examples 2-8 page 12, line 21 -page 14, line 31; examples 2-8 page 17, line 15 - line 27; examples 2-8 page 29, line 13 -page 30, line 27; examples 2-8 page 29, line 13 -page 30; claims 1-15,17,20-23; examples 2-8	1-60
A	US 5 649 904 A (GIANNI ALESSANDRO M) 22 July 1997 (1997-07-22) the whole document	1-60

INTERNATIONAL SEARCH REPORT

International application No.
PCT/EP 99/ 10470

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. [X] Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:

Remark: Although claim59, 60 are directed to a method of treatment of the the human/animal body, the search has been carried out and based on the alleged effects of the compound composition.

2. [ ] Claims Nos.:
because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:

3. [ ] Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. [ ] As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.

2. [ ] As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.

3. [ ] As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:

4. [ ] No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

[ ] The additional search fees were accompanied by the applicant's protest.

[ ] No protest accompanied the payment of additional search fees.

INTERNATIONAL SEARCH REPORT

information on patent family members

Inter[redacted] Application No
PCT/EP 99/10470

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